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New species, transferences and taxonomic notes on American Lamiinae (Coleoptera, Cerambycidae)

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Abstract. – The morphological features of *Asemolea* Bates, 1881 (Calliini) are discussed and expanded, and *Callia ambigua* Bates, 1885 is transferred to *Asemolea*. Additional morphological features are provided for *Murupi* Martins & Galileo, 1998; *M. linearis* Martins & Galileo, 1998 is newly recorded from French Guiana and chromatic variation is reported. Six new species are described: *Scythropopsis tysoni* sp. nov., from Guatemala; *Asemolea rosea* sp. nov. from Guatemala; *Inermestola boliviensis* sp. nov., from Bolivia; *Estola linearis* sp. nov., from Ecuador; *Estoloides elongata* sp. nov., from Panama; and *Eranina monnei* sp. nov., from Panama. The inclusion of *Inermestola chiapasensis* Galileo & Moysés, 2013 in this genus is questioned, indicating the need of study of the species to correct the generic allocation. *Estola compacta* Breuning, 1940 is recorded from Ecuador and Peru, and *E. m-flava* Breuning, 1940 is recorded from Trinidad and Tobago, from the Brazilian states of Amapá and Pará, and a forgotten record from the Brazilian state of Goiás is reported. The second formally known specimen of *Eupogonius fuscovittatus* Breuning, 1974 is illustrated, and a new department record in Guatemala is provided. *Adesmus stephanus* Aurivillius, 1900 is transferred to *Eranina* Monné, 2005, and *Laraesima ochreoapicalis* Breuning, 1973 (Composomatini) is transferred to *Estolomimus* Breuning, 1940 (Desmiphorini). *Laraesima fuliginea* Bates, 1885 is redescribed.

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Introduction

Acanthoderini Thomson, 1860 is a large tribe of Lamiinae. According to Tavakilian & Chevillotte (2021) and Roguet (2022), it includes 67 genera distributed worldwide. *Scythropopsis* Thomson, 1864 is an American genus with 14 species distributed from Mexico to southern South America (Monné 2022). The material examined allowed us to describe a new species from Guatemala.

Calliini Thomson, 1864 includes 45 genera of which only *Neocallia* Fisher, 1933 does not occur in America (Tavakilian & Chevillotte 2021). *Asemolea* Bates, 1881 includes six species distributed from Mexico to northern South America (Monné 2022). The definition of the genus is updated, a new species is described, and another species transferred from *Callia* Audinet-Serville, 1835 to *Asemolea*.

Composomatini Thomson, 1857 includes 13 genera distributed from Mexico to southern South America. *Laraesima* Thomson, 1868 was reviewed by Monné (1980b). Even so, the genus is still problematic and, apparently, encompasses species of more than one genus. Here we redescribe *L. fuliginea* (Bates, 1865), a species not examined by Monné (1980b), which probably

belongs to a different genus, and a new Guatemalan department record is provided for the species.

Currently, Desmiphorini Thomson, 1860 includes 529 American species distributed in 85 genera (Tavakilian & Chevillotte 2021; Bezark 2022); of these, 42 genera include a single known species, and only five include more than 20 species. Here we describe three new species, question the inclusion of *Inermestola chiapasensis* Galileo & Moysés, 2013 in this genus, provide new records, illustrate *Eupogonius fuscovittatus* Breuning, 1974, and transfer *Laraesima ochreoapicalis* Breuning, 1973 to *Estolomimus* Breuning, 1940.

Hemilophini Thomson, 1868 is a large American tribe of Lamiinae, including 135 genera distributed from the United States of America to southern South America (Roguet 2022). Here we describe a new species of *Eranina* Monné, 2005, transfer *Adesmus stephanus* Aurivillius, 1900 to *Eranina*, and illustrate *Eupogonius fuscovittatus* Breuning, 1974.

Material and Methods

Photographs were taken in the MZSP with a Canon EOS Rebel T7i DSLR camera, Canon MP-E 65mm f/2.8 1-5X macro lens, controlled by Zerene Stacker AutoMontage software. Measurements were taken in “mm” using measuring ocular

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Hensoldt/Wetzlar - Mess 10 in the Leica MZ6 stereomicroscope, also used in the study of the specimens.

The species were identified using original descriptions, redescriptions, photographs of the holotypes, and comparisons with specimens of the MZSP collection.

The collection acronyms used in the text are as follows:

- CASC: California Academy of Sciences, San Francisco, California, USA.
- FSCA: Florida State Collection of Arthropods, Gainesville, Florida, USA.
- FWSC: Fred W. Skillman, Jr. collection, Phoenix, Arizona, USA.
- LGBC: Larry G. Bezark collection, Sacramento, California, USA.
- MZSP: Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil.
- WHTC: William H. Tyson Collection, Coarsegold, California, USA.

Results

ACANTHODERINI Thomson, 1860

Scythropopsis tysoni sp. nov.

(Fig. 1-8)

ZooBank: <http://zoobank.org/D91701BC-6A84-4DD2-9945-31565432082B>

Holotype, ♂, GUATEMALA, *Quetzaltenango*: Fuentes Georginas, 2400 m, 14-16.VI.2015, W.H. Tyson leg. (CASC, formerly WHTC).

Paratype, ♀, same data as holotype (WHTC).

Description of the holotype (Fig. 1-4).

Coloration. – Integument mostly black; ventral mouthparts dark reddish brown, except palpomeres mostly black; pedicel, basal half of antennomeres III–IV reddish brown; basal half of antennomere V dark reddish brown, femoral peduncles, and wide central area of tibiae dark reddish brown.

Head. – Frons, vertex, and area behind upper eye lobes very finely, densely punctate, except smooth median groove; with bristly, white and yellowish brown pubescence interspersed, and long, erect, moderately abundant brownish setae interspersed. Area behind lower eye lobes and between eyes and antennal tubercles with dense, white and yellowish brown pubescence close to eye, glabrous close to prothorax; with long, erect white setae interspersed with pubescence behind lower eye lobes. Genae with dense, white and yellowish brown pubescence close to eye, sparser on remaining surface, almost glabrous on apex; with long, erect, sparse brownish setae interspersed. Wide central area of postclypeus with bristly, dense, white and yellowish brown pubescence close to frons, and long, abundant, white and yellowish brown setae directed forward close to anteclypeus; with long, erect, sparse brownish setae interspersed. Sides of postclypeus glabrous. Labrum convex, coplanar with anteclypeus at posterior 2/3, inclined at anterior third; with moderately short and abundant, both white and yellowish brown setae on posterior 2/3, glabrous on inclined area, and fringe of short yellow setae on anterior margin; with long, erect brownish setae interspersed on coplanar area close to inclined area. Gula mentum smooth, glabrous, except narrow anterior area slightly depressed, transversely striate-punctate, with short, bristly, moderately sparse, white and yellowish brown setae, and a few long, erect brownish setae interspersed. Eye lobes united by single row of irregular ommatidia; distance between upper eye lobes 0.44 times distance between outer margins of eyes; in frontal view, distance between lower eye lobes 0.65 times distance between outer margins of eyes. Antennae 1.65 times elytral length, reaching elytral apex at base of antennomere VIII. Scape somewhat pedunculate-clavate; with white and yellowish-brown pubescence not obscuring integument, and long, erect, white and brownish setae interspersed, erect setae more abundant dorsally. Pedicel with white pubescence basally, mostly brownish on remaining surface; with a few long, white and brownish setae interspersed ventrally. Antennomeres III–IV cylindrical on basal half, slightly, gradually widened toward apex on posterior half; with white pubescence basally, yellowish brown with white short setae interspersed on remaining basal half, and blackish pubescence on posterior half; with long, erect dark setae ventrally, distinctly more abundant on posterior third. Antennomeres V–XI with white pubescence on anterior region, about basal half on V–VIII, about basal third on IX–XI; remaining surface with blackish pubescence; antennomeres V–X with long, erect dark setae ventrally.

Antennal formula based on length of antennomere III:

– Scape = 0.70. – Pedicel = 0.21. – IV = 0.89. – V = 0.54. – VI = 0.40. – VII = 0.38. – VIII = 0.27. – IX = 0.21. – X = 0.19. – XI = 0.16.

Thorax. – Prothorax wider than long; laterally with large, conical tubercle with blunt apex centrally. Sides of prothorax moderately finely, sparsely punctate; with abundant, white and yellowish brown pubescence not obscuring integument, and long, erect, moderately abundantly white setae interspersed. Pronotum with long, arched carina on each side, from anterior quarter to about posterior third, and longitudinal carina centrally, from near anterior margin to about posterior third; coarsely, moderately sparsely punctate, punctures distinctly sparser between carinae; with abundant, bristly, white and yellowish brown pubescence, absent on carinae; with long, erect dark setae interspersed, more abundant laterally. Sides of prosternum with abundant, bristly, white and yellowish brown pubescence not obscuring integument, and long, erect, moderately abundant white setae interspersed; central area with sparse white pubescence and sparse yellowish brown pubescence interspersed (yellowish brown pubescence more abundant on anterior quarter), and erect, sparse white setae interspersed. Prosternal process with sparse white pubescence and long, erect setae of same color interspersed, erect setae more abundant apically; narrowest area 0.7 times procoxal width. Sides of ventral surface of meso- and metathorax with dense, white and yellowish brown pubescence, and long, erect, abundant white setae interspersed; central area with white pubescence not obscuring integument, except glabrous posteroventral area of metaventre, and long, erect, moderately abundant white setae interspersed. Mesoventral process 1.15 times mesocoxal width. Scutellum with abundant, white and yellowish brown pubescence on anteroventral area, slightly sparser, mostly yellowish brown on remaining surface.

Elytra. – With moderately elevated and obliquely arched carina on inner dorsobasal quarter, with small tubercles dorsally, following slightly elevated, subparallel-sided to suture toward posterior fifth; another carina on outer dorsal surface, from about basal quarter, fused apically with the former; coarsely, moderately abundantly punctate on basal third, punctures finer, sparser toward apex; with abundant white pubescence partially obscuring integument, with somewhat irregular, longitudinal rows of yellowish brown pubescence interspersed, except oblique, moderately narrow brown pubescent band on basal quarter of dorsal surface, starting before lateral curvature and ending at apex of arched carina, and wide, oblique brown pubescent band starting on middle of dorsal surface, not reaching lateral curvature and suture; with long, erect, moderately abundant brownish setae interspersed throughout; apex concave centrally.

Legs. – Femoral peduncles, base of femoral clubs with abundant white pubescence not obscuring integument, and long, erect, abundant white setae interspersed ventrally; remaining surface of femoral clubs and tibiae with abundant, white and yellowish brown pubescence not obscuring integument, white pubescence more abundant on meso- and metatibiae, except dorsal sulcus of mesotibiae with thick, abundant dark setae; tibiae with long, erect white, yellowish brown and dark setae interspersed, white setae more abundant on meso- and metatibiae. Metatarsomere I slightly longer than II–III together.

Abdomen. – Sides of ventrites with abundant yellowish brown pubescence, and white pubescence and long, erect, abundant white setae interspersed; central area with white pubescence not obscuring integument, sparser on ventrite 5, and yellowish brown pubescence interspersed; ventrite 5 1.5 times length of ventrite 4, with apex widely truncate.

Female (Fig. 5-8). – Differs from male by the shorter antennae, 1.4 times elytral length, reaching elytral apex at base of antennomere XI, by ventrite 5 three times longer than ventrite 4, and distinctly narrower apically, and also appears to differ in having a more pronounced carina along the elytral suture medially.

Dimensions (mm) (holotype ♂/paratype ♀).

- Total length, 7.05/9.95
- Prothoracic length, 1.40/1.80
- Anterior prothoracic width, 1.85/2.45
- Posterior prothoracic width, 1.80/2.35
- Maximum prothoracic width, 2.30/2.85
- Humeral width, 2.60/3.80
- Elytral length, 4.80/6.85.



Fig. 1-8. *Scythropopsis tysoni* sp. nov.

1-4. Holotype ♂. 1. Dorsal habitus. 2. Ventral habitus. 3. Lateral habitus. 4. Head, frontal view.
5-8. Paratype ♀. 5. Lateral habitus. 6. Dorsal habitus. 7. Ventral habitus. 8. Head, frontal view.

Etymology. – This species is named for William H. Tyson, Coarsegold, California, USA, friend and colleague of the first author, who collected the holotype and paratype.

Remarks. – *Scythropopsis tysoni* sp. nov. is somewhat similar to *S. intricata* Santos-Silva, Botero & Wappes, 2020 (see photographs of the holotype on Bezark 2022), but differs as follows:

- (i) frons with white and yellowish brown pubescence.
- (ii) distance between upper eye lobes 0.5 times length of the antennomere IV.
- (iii) elytra slender and not strongly narrowed toward apex.
- (iv) ventral surface of the body with long, erect and abundant white setae.

In *S. intricata*:

- (i) the frons has dense white pubescence.
- (ii) distance between upper eye lobes slightly shorter than length of the antennomere IV.
- (iii) elytra are wider and strongly narrowed toward apex.
- (iv) ventral surface of the body without long and abundant white setae.

It also differs from *S. sallei* (Thomson, 1865) by:

- (i) the elytra not strongly narrowed toward apex.
- (ii) shorter distance between upper eye lobes (elytral strongly narrowed toward apex and distance between upper eye lobes slightly shorter than length of antennomere IV in *S. sallei*).

CALLIINI Thomson, 1864

Asemolea Bates, 1881

Asemolea Bates, 1881: 194.

Aurivillius, 1923: 601 (cat.).

Galileo & Martins, 1991: 244, 248.

Monné, 1995: 79; 2005: 315 (cat.); 2012: 88.

Santos-Silva *et al.*, 2019: 9 (key).

Ramírez Hernández *et al.*, 2019: 91 (key).

Monné, 2022: 539 (cat.).

Remarks. – Bates (1881) described *Asemolea* as follows (translated): “Genus near *Drycothaea*, it differs especially by the thorax unarmed laterally, rounded. Body short, subcylindrical, with long and dense erect setae. Head inserted [retractile]; frons short; genae obtuse, narrowed. Thorax short, sides rounded centrally. Elytra linearly punctate, apex obtuse. Mesoventral process slightly convex. Tarsal claws with a single tooth internally [claws apendiculate]. Antennae shorter than body, stout, with apex acute and with long setae ventrally.” After the original description only Galileo & Martins (1991) provide some additional information about the features of the genus. According to them, in the key separating *Asemolea* from *Euryestola* Breuning, 1940 (translated): “Mesotibiae without sulcus on outer surface; upper eye lobe separated from lower eye lobe by a single row of ommatidia,” leading to *Asemolea*; mesotibia with sulcus on outer surface; upper eye lobe separated from lower eye lobe by more than one row of ommatidia,” leading to *Euryestola*. In fact, the sulcus on the outer surface of the mesotibiae in *Euryestola* is the dorsal sulcus, which is distinctly more apical and reaches the sides of the tibiae in this genus, while it is placed more closely towards middle and not distinctly reaching the sides in *Asemolea*.

The information on the number of rows of ommatidia between the eyes lobes, apparently, was based on two specimens of *A. setosa* Bates, 1881, belonging to the MZSP collection. It was not possible to be sure if they really belong to *A. setosa*, although they belong in *Asemolea*, because the pubescent color is different (most grayish white in the specimens from the

MZSP collection, mostly yellowish brown in the lectotype and paralectotypes) and the ommatidia appear to be coarser than in the paralectotype illustrated in MCZbase (2021). However, the area of connection between the eye lobes in the specimens from the MZSP collection has two rows of ommatidia (Fig. 10) in the male, and one partial row in the female (Fig. 9). The paralectotype of *A. setosa* has two distinct rows of ommatidia (see MCZbase 2021). In the description of *A. macaranduba* Galileo & Martins (1998) and *A. flava* Martins & Galileo, 2006 the authors already had reported that the eye lobes are connected by three rows of ommatidia. However, both Galileo & Martins (1998) and Martins & Galileo (2006) did not comment on the problem with the concept of the genus in relation to this feature. Even so, the key to genera of Calliini by Galileo & Martins (1991) allows separating *Asemolea* from the other genera, excluding the information on the eyes in the alternative of couplet “6.”

In addition to the features pointed out by Bates (1881), we found the following features of *Asemolea*:

- (i) General appearance not “lampyroid”.
- (ii) Eye lobes interconnected by one (usually only on part of the area) (Fig. 9) or more rows of ommatidia (Fig. 10 & 16).
- (iii) Basal flagellomeres without dense setae ventrally.
- (iv) Flagellomeres not flattened.
- (v) Scape without apical cicatrix.
- (vi) Prothorax unarmed or with minute tubercle centrally.
- (vii) Mesoventrite without distinct tubercle (at most, longitudinal slightly carinate).
- (viii) Dorsal sulcus of mesotibiae placed closer to middle than apex.
- (ix) Metatarsomere I not distinctly longer than II and III together.

Asemolea ambigua Bates, 1885 **comb. nov.**

(Fig. 11–12)

Callia (Callia) ambigua Bates, 1885: 424.

Aurivillius, 1923: 600 (cat.).

Gilmour, 1965: 648 (cat.).

Callia ambigua

Blackwelder, 1946: 626 (checklist).

Chemsak & Linsley, 1970: 408 (lect.).

Galileo & Martins, 1991: 260.

Chemsak *et al.*, 1992: 161 (cat.).

Monné & Giesbert, 1994: 296 (checklist).

Monné, 1995: 92 (cat.).

Noguera & Chemsak, 1996: 408 (checklist).

Galileo & Martins, 2002: 51 (key).

Monné, 2005: 316 (cat.).

Monné & Hovore, 2006: 230 (checklist).

Monné, 2022: 540 (cat.).

Remarks. – Bates (1885) described *Callia ambigua* as follows (translated): “Narrowly elongated, slightly widened posteriorly, with erect setae, black, grayish pubescent; vertex and lateral bands on the prothorax densely golden pubescent; elytra entirely black, or with short humeral band (narrowed posteriorly) fulvous, or the anterior half (except suture and epipleural margin) fulvous; antennomeres III–XI grayish basally; prothorax short, cylindrical, barely tuberculate centrally on sides; elytra striate-punctate; longitudinally depressed along most of sutural region.” From what it is possible to see in the photographs of the lectotype, the sides of the prothorax have no tubercle. The same condition was found in the specimen examined by us. Probably, in some species, the central area of the sides have the pubescence somewhat bristly,

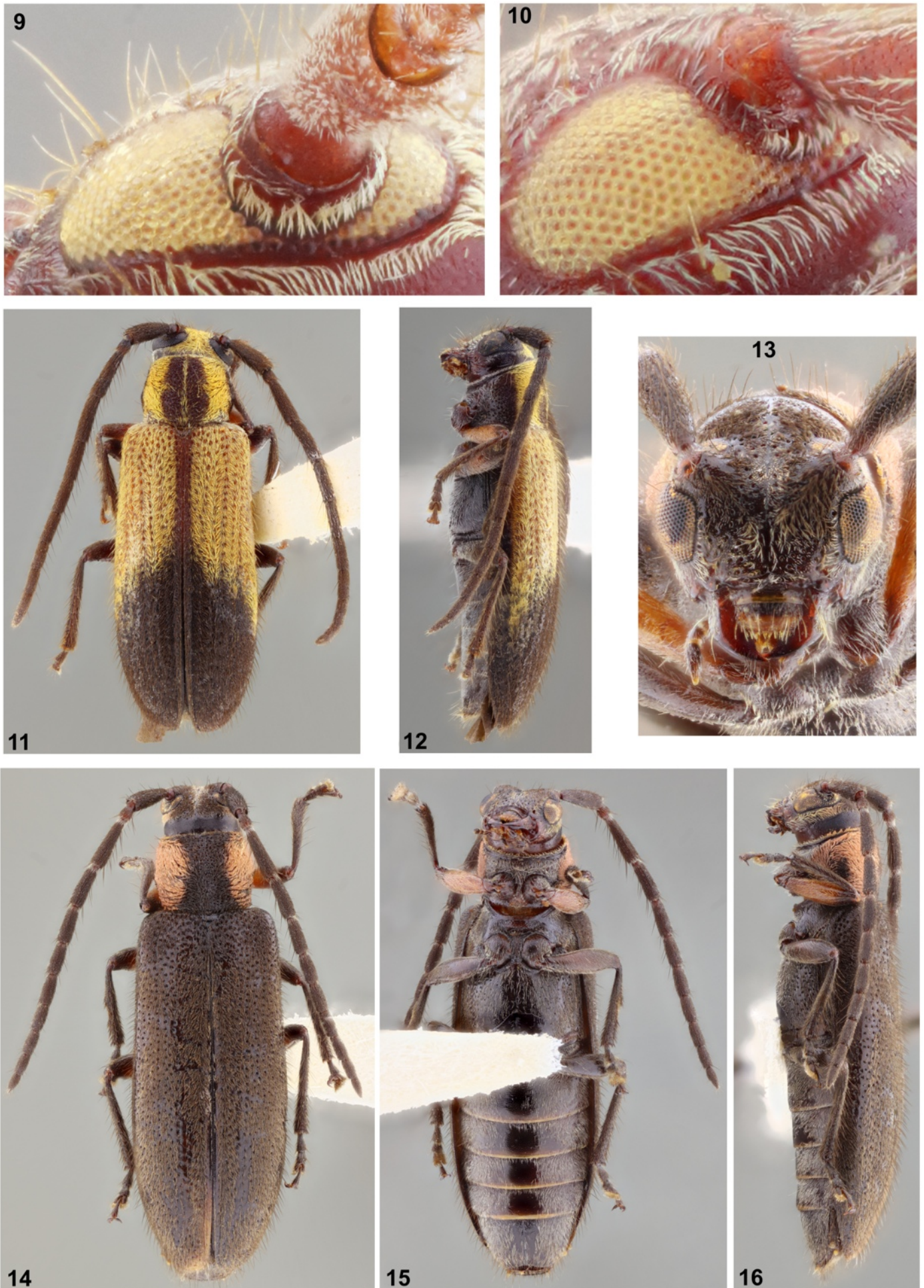


Fig. 9-16. *Asemolea* spp.

9-10. *A. setosa* Bates, 1881, eye. 9. ♀. 10. ♂.

11-12. *A. ambigua* (Bates, 1885), ♀. 11. Dorsal habitus. 12. Lateral habitus.

13-16. *A. rosea* sp. nov., holotype ♀. 13. Head, frontal view. 14. Dorsal habitus. 15. Ventral habitus. 16. Lateral habitus.

giving the appearance of a small tubercle. The same occurs in the new species of *Asemolea* described here. As the lateral tubercles in *Callia* Audinet-Serville, 1835 are very conspicuous, we transfer *Callia ambigua* to *Asemolea*. Furthermore, according to Bates (1885), “This species would fit almost equally well the genus *Asemolea*,” which already suggested his doubt about the genus of the species.

Material examined. – MEXICO, *Veracruz*: Lake Catemaco, 1 ♀, 1-2.V.1969, Bright & Campbell leg. (MZSP).

***Asemolea rosea* sp. nov.**

(Fig. 13-16)

ZooBank: <http://zoobank.org/2CD06711-01E4-471E-8619-17EB0506FBF0>

Holotype, ♀, GUATEMALA, *Alta Verapaz*: 32 mi SE Coban, 6000', 26.VII.1974, O'Brien & Marshall leg. (CASC, formerly LGBC).

Description of the holotype

Coloration. – Integument mostly dark brown, almost black; ventral mouthparts dark reddish brown, except palpomeres black with apex yellowish brown; anterior area of anteclypeus and labrum dark yellowish brown; profemora dark orangish brown except dark brown apex; mesofemora brown except dark brown apex; apex of ventrites 1–4 yellowish brown.

Head. – Frons moderately coarsely and abundantly punctate, except smooth area close to median groove; with oblique, wide, dense brownish pubescent band on each side, from median groove to base of antennal tubercle, bristly pale yellow setae on each side of median groove, from middle to near clypeus, gradually sparser toward clypeus, yellowish white pubescence close to eyes, minute, decumbent, sparse yellowish white setae on remaining surface of anterior half, minute, decumbent, moderately abundant yellowish setae between brownish pubescent band, and long, erect, sparse dark brown setae interspersed throughout. Central area between antennal tubercles and upper eye lobes moderately coarsely, sparsely punctate; with abundant yellowish white pubescence partially obscuring integument, and sides with dense brown pubescence; with long, erect, sparse dark brown setae interspersed laterally. Remaining surface of vertex moderately coarsely, sparsely punctate, except smooth area close to prothorax; with dense brown pubescence obscuring integument, except glabrous longitudinal central area and area close to prothorax, and narrow yellowish white pubescent band close to glabrous area; with a few long, erect dark brown setae laterally. Area behind eyes, moderately coarsely and sparsely punctate close to superior area of upper lobe, finely, densely punctate on remaining surface close to eye, smooth close to prothorax; with dense brown pubescence on superior area close to upper lobe, dense yellowish brown pubescence on remaining surface close to eye, except yellowish white pubescence close to genae, glabrous close to prothorax; with a few long, brownish setae interspersed on pubescent area. Genae slightly rugose-punctate, except smooth apex; with yellowish white pubescence not obscuring integument, denser toward ventral surface, except glabrous apex. Antennal tubercles finely, sparsely punctate; with sparse brown pubescence and long, erect, dark brown setae interspersed. Wide central area of postclypeus with bristly yellowish white pubescence not obscuring integument, longer and denser centrally; with long, erect, sparse brownish setae and one long, erect seta on each side, dark brown basally, gradually yellowish toward apex (longer than remaining erect setae). Labrum coplanar with anteclypeus at posterior third, inclined at anterior 2/3; finely, abundantly punctate on coplanar area, nearly smooth on remaining surface; with long, erect yellowish setae on coplanar area and sides of inclined area; anterior margin with dense fringe of golden setae. Gulamentum smooth glabrous, except narrow anterior area with sparse yellowish white pubescence and long, erect brownish setae interspersed. Distance between upper eye lobes 0.32 times distance between outer margins of eyes; in frontal view, distance between lower eye lobes 0.61 times distance between outer margins of eyes. Antennae slightly shorter than elytra, reaching

posterior third of elytra; with abundant brownish pubescence not obscuring integument, except a few grayish white setae on dorsal apex of scape, sparse grayish white pubescence on narrow apex of pedicel, and grayish white pubescence on base of antennomeres III–VII, distinctly sparser toward VII; scape and pedicel with erect, sparse dark brown setae interspersed throughout, longer ventrally on pedicel; antennomeres III–XI with dark brown setae ventrally, longer, moderately abundant on III, gradually shorter and sparser toward XI; antennomeres III–VI with short, erect, sparse dark brown setae interspersed dorsally; dorsal apex of antennomeres III–X with dark brown setae, gradually sparser toward X; apex of antennomere XI with moderately long yellowish setae.

Antennal formula based on length of antennomere III:

– Scape = 0.89. – Pedicel = 0.26. – IV = 0.81. – V = 0.73. – VI = 0.58. – VII = 0.58. – VIII = 0.52. – IX = 0.47. – X = 0.45. – XI = 0.45.

Thorax. – Prothorax wider than long; sides divergent from anterolateral angles to middle, convergent from this area to posterolateral angles, with minute tubercle centrally. Sides of prothorax with sculpturing as on pronotum; with dense salmon-colored pubescence, except dark brown pubescence on wide longitudinal band near pronotum, from apex of anterior third to posterior margin, and glabrous narrow area close to anterior margin. Pronotum coarsely, abundantly punctate; with dense salmon-colored pubescence laterally (lighter depending on light intensity), inner margin of this area arched, with abundant dark brown pubescence on remaining surface not obscuring integument, and long, erect, both brown and yellowish setae interspersed. Prosternum coarsely, sparsely punctate, except somewhat rugose narrow anterior area; with abundant yellowish white pubescence not obscuring integument, sparser close to anterior margin. Prosternal process with bristly yellowish white pubescence not obscuring integument; narrowest area 0.15 times procoxal width. Mesoventrite slightly depressed on wide central area; with sparse yellowish white pubescence, almost absent on antero-central region. Mesanepisternum and mesepimeron with brownish pubescence, denser on mesepimeron. Mesoventral process strongly inclined close to mesoventrite, moderately longitudinally sulcate centrally, and slightly widened apically; apex 0.38 times mesocoxal width. Metanepisternum with abundant brown pubescence not obscuring integument. Sides of metaventricle coarsely punctate, punctures denser close to metanepisternum, almost smooth centrally; with abundant brown pubescence not obscuring integument laterally, and sparser yellowish white pubescence centrally. Scutellum with sparse brown pubescence.

Elytra. – Coarsely, abundantly punctate on basal third, punctures slightly, gradually finer and sparser toward apex; apex slightly, individually rounded; with abundant brown pubescence not obscuring integument, except yellowish brown pubescence on lateral curvature, and abundant, erect dark brown setae interspersed.

Legs. – Profemora with whitish pubescence not obscuring integument; meso- and metafemora with light yellowish brown pubescence not obscuring integument dorsally and laterally, yellowish, bristly ventrally. Protibiae with yellowish brown pubescence not obscuring integument, except dense, bristly brown pubescence on posterior 2/3 of ventral surface, and long, erect brown setae interspersed dorsally and laterally; mesotibiae with brown pubescence not obscuring integument, and suberect, thick, moderately short dark brown setae on dorsal surface, before and after dorsal sulcus, and on posterior third of ventral surface, and bristly yellowish brown pubescence on dorsal sulcus; metatibiae with brownish pubescence not obscuring integument, except thick, suberect dark brown setae on posterior third, and long, erect yellowish brown setae interspersed. Metatarsomere I slightly shorter than II–III together.

Abdomen. – Ventrites with grayish yellow pubescence laterally, partially obscuring integument, almost glabrous centrally, except narrow pubescent band basal area of ventrites 1–4, sparse pubescence on central basal 2/3 of ventrite 5, and moderately abundantly pubescence on posterior third of ventrite 5; with long, erect, sparse yellowish setae interspersed on sides. Ventrite 5 with narrow sulcus centrally on basal third, and triangularly depressed on central area of posterior 2/3; apex of ventrite 5 centrally emarginate.



Fig. 17-21. *Laraesima fuliginea* Bates, 1885, ♀.

17. Dorsal habitus. 18. Ventral habitus. 19. Lateral habitus. 20. Head, frontal view. 21. Oblique view.

Dimensions (mm).

- Total length, 7.80
- Prothoracic length, 1.15
- Anterior prothoracic width, 1.30
- Posterior prothoracic width, 1.40
- Maximum prothoracic width, 1.55
- Humeral width, 2.05
- Elytral length, 5.60.

Etymology. – The species epithet *rosea* (Latin) refers to the salmon-colored pubescence, unique among species in this genus.

Remarks. – *Asemolea rosea* sp. nov. differs from all other species of the genus by the sides of the pronotum and most sides of the prothorax covered with dense salmon-colored pubescence, which is absent in the other species. *Asemolea ambigua* (Fig. 11-12) has dense pubescence on the same areas, but it is yellowish brown, and the vertex is also covered by dense yellowish brown pubescence (mostly brown in *A. rosea*), and the elytral punctures are coarser (finer in *A. rosea*).

COMPSOSOMATINI Thomson, 1857***Laraesima fuliginea*** (Bates, 1885)

(Fig. 17-21)

Pennesada fuliginea Bates, 1885: 361.

Aurivillius, 1923: 339 (cat.).

Blackwelder, 1946: 602 (checklist).

Chemsak & Linsley, 1970: 415 (lect.).

Laraesima fuliginea

Breuning, 1961: 290 (cat.).

Monné, 1980b: 234.

Breuning, 1980: 15.

Chemsak *et al.*, 1992: 123 (cat.).

Monné & Giesbert, 1994: 213 (checklist).

Monné, 1994: 19 (cat.).

2005: 366 (cat.); Hovore, 2006: 376 (distr.).

Monné, 2022: 620 (cat.).

Material examined. – GUATEMALA, *Baja Verapaz* (new department record): 3 km S Purulha, 1650-1800, montane forest, 15°12.965'N, 90°13.142'W, 1 ♂, 27-30.VI.2012, E. Fuller leg. (WHTC).

Redescription of the female

Coloration. – Integument mostly dark brown; ventral mouthparts reddish brown, except pale yellow apex of palpomeres; anteclypeus mostly brownish; anterior area of labrum yellowish brown; scape dark brown on basal half, brownish on posterior half; pedicel reddish brown; antennomeres III–X pale yellow basally (missing antennomeres XI on right antenna, and antennomeres X–XI on left antenna), light reddish brown on remaining surface. Trochanters, base and apex of femora, and approximately basal third of tibiae dark reddish brown; tarsi mostly yellowish brown.

Head. – Frons coarsely, abundantly punctate; triangularly concave from middle to area between antennal tubercles; with abundant brown pubescence not obscuring integument, except small grayish white pubescent macula on each side of middle, and sparse grayish white pubescence interspersed close to eyes; with short, erect, sparse yellowish brown setae interspersed. Area between antennal tubercles and upper eye lobes with sculpturing as on frons; with brownish pubescence distinctly sparser than on frons, except grayish white pubescence close to posterior area of base of antennal tubercles; vertex distinctly, moderately abruptly elevated after upper eye lobes, coarsely, sparsely punctate, punctures slightly sparser close to prothorax; with sparse brown pubescence. Area behind upper eye lobes coarsely, moderately abundantly punctate; with brown pubescence not obscuring integument. Area behind lower eye lobes coarsely, partially confluent punctate (punctures coarser than behind upper eye lobes); with brown pubescence not obscuring integument, except grayish white

pubescence close to eye. Genae coarsely, sparsely punctate, except smooth apex; with grayish white pubescence not obscuring integument, except glabrous smooth area. Antennal tubercles with sculpturing as on frons; with brown pubescence not obscuring integument. Wide central area of postclypeus transversely carinate centrally; with sparse brown pubescence close to frons, short, sparse brownish setae, and long, erect setae of same color interspersed close to anteclypeus. Sides of postclypeus glabrous. Labrum coplanar with anteclypeus on posterior 3/4, inclined at anterior quarter; coplanar area finely, somewhat rugose-punctate, especially laterally, with short, decumbent, sparse, both yellowish brown and grayish white setae, and long, erect, yellowish brown setae interspersed; anterior margin with dense fringe of golden setae. Eye lobes united by single row of ommatidia; distance between upper eye lobes 0.23 times distance between outer margins of eyes; in frontal view, distance between lower eye lobes 0.56 times distance between outer margins of eyes. Antennae 1.4 times elytral length (from base of scape to apex of antennomere X), reaching elytral apex at posterior third of antennomere X. Scape strongly widened after base, subcylindrical on posterior half; with abundant brown pubescence not obscuring integument dorsally and laterally, except narrow white pubescent band on apex of dorsal surface, moderately sparse white pubescent on posterior third of sides and entire ventral surface; with a few short, suberect yellowish brown setae interspersed. Pedicel with yellowish pubescence not obscuring integument dorsally, except white pubescence on apex, sparse yellowish pubescence on basal half of sides and ventral surface, sparse white pubescence on remaining surface, and long, erect yellowish brown setae ventrally. Antennomeres III–X with abundant yellowish brown pubescence not obscuring integument, except white pubescence on base and fringe of white pubescence on apex; antennomere III with long, erect, moderately abundant yellowish brown setae ventrally, longer, denser, directed backward on apex; antennomere IV arched, with short, erect yellowish brown setae ventrally, longer, denser, directed backward apically; antennomeres V–VI with a few short yellowish brown setae apically.

Antennal formula based on length of antennomere III:

– Scape = 1.04. – Pedicel = 0.24. – IV = 1.52. – V = 0.82. – VI = 0.69. – VII = 0.69. – VIII = 0.65. – IX = 0.61. – X = 0.56.

Thorax. – Prothorax wider than long; laterally with conical tubercle centrally. Sides of prothorax coarsely, moderately abundantly punctate; with abundant yellowish brown pubescence not obscuring integument. Pronotum with three elevated tubercles, one on each side of anterior third, another less elevated than anterior ones, placed centrally slightly beyond middle; coarsely, abundantly punctate; with brown pubescence on anterior 2/3 not obscuring integument, yellowish brown pubescence on posterior third not obscuring integument, except narrow band of yellow pubescence centrally close to anterior margin, yellow pubescent band centrally close to posterior margin, and four white pubescent maculae centrally, the two innermost distinctly sparser. Prosternum coarsely, moderately sparsely punctate; with abundant yellowish brown pubescence laterally, not obscuring integument, and shorter, sparser pubescence of same color centrally. Prosternal process coarsely, abundantly punctate; with grayish white pubescence laterally, and sparser, shorter yellowish pubescence centrally; narrowest area 0.23 times procoxal width. Ventral surface of meso- and metathorax coarsely, moderately abundantly punctate; mesothorax mostly with light yellowish brown pubescence not obscuring integument; metanepisternum with grayish white pubescence not obscuring integument, except light yellowish brown pubescence on posterior area; metaventricle with grayish white pubescence not obscuring integument. Scutellum with dense yellowish brown pubescence.

Elytra. – With moderately elevated, longitudinal centrobasal crest on anterior fifth, and four longitudinal carinae not reaching apex (outer most carina only reaching apex of anterior third); coarsely, moderately abundantly punctate on basal half, punctures gradually finer and sparser toward apex; with abundant brownish pubescence not obscuring integument close to suture on anterior 3/4, dark brown pubescence not obscuring integument on remaining surface, and abundant, irregular grayish white pubescent maculae throughout (grayish white maculae more abundant on posterior quarter); apex rounded, subtruncate near sutural angle.

Legs. – Femora mostly with whitish pubescence, except area near apex of dorsal surface with yellowish brown pubescence. Tibiae with brownish pubescence not obscuring integument and whitish pubescence interspersed on some areas, except bristly, mostly dark pubescence on posterior third of ventral surface of protibiae and entire surface of meso- and metatibiae. Metatarsomere I slightly shorter than II–III together.



Fig. 22-26. *Murupi linearis* Martins & Galileo, 1998.

22-25. ♂, French Guiana. 22. Dorsal habitus. 23. Ventral habitus. 24. Lateral habitus. 25. Head, frontal view.
26. ♂, Brazil (Amazonas), dorsal habitus.

Abdomen. – Ventrites 1-4 moderately coarsely and sparsely punctate anteriorly, smooth posteriorly; ventrite 5 moderately coarsely and sparsely punctate throughout; with grayish white pubescence not obscuring integument.

Dimensions (1 ♀) (mm)

- Total length, 7.60
- Prothoracic length, 1.55
- Anterior prothoracic width, 1.55
- Posterior prothoracic width, 1.70
- Maximum prothoracic width, 2.00
- Humeral width, 2.70
- Elytral length, 5.45.

Remarks. – Following the key by Monné (1980a), *Laraesima fuliginea* would belong to *Parapythais* Monné, 1980 (translated):

“Elytra without erect setae,” leading to *Parapythais*; “Elytra with erect setae,” leading to the other genera of Composomatini.

However, *L. fuliginea* is distinctly different from *P. melzeri* Monné, 1980, the unique species of the genus, which has:

- (i) the upper eye lobes separated by a distance equal to four times the width of one upper lobe (separated by a distance slightly wider than the width of one upper lobe in *L. fuliginea*).
- (ii) antennomeres III and IV with similar length (IV distinctly longer than III in *L. fuliginea*).
- (iii) humeral angles strongly projected (not projected in *L. fuliginea*).
- (iv) elytra triangular shaped (not so in *L. fuliginea*).

Laraesima fuliginea was not included in the key to species of the genus from Monné (1980b). However, this key really allows to include *L. fuliginea* in the genus.

Apparently, *Laraesima* includes species belonging to three genera:

- (i) elytra without erect setae.
- (ii) elytra with long and slender dark setae.
- (iii) elytra with very short, thick white setae.

Laraesima fuliginea was described based on at least one male and one female from Guatemala (Cerro Zunil and Panajachel). According to Selander & Vaurie (1962), Cerro Zunil is in the department of Quetzaltenango and Panajachel is in the department of Sololá. Chemsak & Linsley (1970) designated as lectotype a specimen from Panajachel. However, Monné (2022) mistakenly reported “Guatemala, Quezaltenango: Cerro Zunil” as the type locality.

The only difference found between the specimen examined and the lectotype is the elytra with grayish white pubescent maculae. According to Bates (1885), the pubescence is fulvous-ochraceous in the specimens examined by him, with agrees well with photographs of the lectotype (see, for example, Bezark 2022). We believe that the difference in the color of the maculae is just specific variation.

DESMIPHORINI Thomson, 1860

Murupi Martins & Galileo, 1998

Murupi Martins & Galileo, 1998: 263.
Monné, 2005: 421 (cat.); 2012: 96; 2022: 707 (cat.).

Remarks. – Martins & Galileo (1998) described *Murupi* as follows (translated): “Elongated general appearance. Body and appendages without setae. Frons transverse. Area between antennal tubercles flat. Eyes coarsely faceted; lower eye lobes longer than three times genal length; upper eye lobes with 5 rows of ommatidia, distance between them larger than width of a lobe. Antennae surpassing the elytral apex. Scape without apical cicatrix, subcylindrical, with half the length of the antennomere III. Antennomere III almost twice length of IV. Prothorax as long as wide; sides regularly rounded. Pronotum convex, without gibbositities. Elytra densely punctate; apex rounded. Prosternal process without tubercle. Mesotibiae sulcate on posterior third. Femora fusiform.” Additionally according to them, *Murupi* differs from *Inermestoloides* Breuning, 1966, *Inermestola* Breuning, 1942, and *Piimuna* Martins & Galileo, 1998 by the very elongate general appearance and by antennomere III being twice the length of the scape and antennomere IV.

To the features presented in the original description, we add the following: prothorax with distinct basal constriction; prothorax, ventral surface of meso- and metathorax, and ventrites densely punctate. The punctation was mentioned in the original description of *M. linearis* Martins & Galileo, 1998; we believe that it is a generic feature. Although it was indicated that the body has no erect setae, there are a few long and erect setae on the frons and pronotum; also, the upper eye lobes may have five or six rows of ommatidia, and the metafemora is more distinctly clavate than fusiform.

As females of this genus remain unknown, the description of *Murupi* is based only on males.

Murupi linearis Martins & Galileo, 1998

(Fig. 22-26)

Murupi linearis Martins & Galileo, 1998: 257.

Monné, 2005: 421 (cat.).

Monné & Hovore, 2006: 249 (checklist).

Monné *et al.*, 2017: 352 (holotype).

Monné, 2022: 707 (cat.).

Remarks. – *Murupi linearis* was described based on a single male from Brazil (Amazonas). Martins & Galileo (1998) described the integument as reddish, darker on the apex of the flagellomeres, the femora, and the tibiae. Seeing photographs of the holotype, this

color description appears to agree well with it. However, the specimens now examined allow indicating that the general color may be distinctly darker. In both specimens examined the head, thorax, most of the elytra and abdomen is dark brown, almost black.

Material examined.

BRAZIL

– Amazonas: Reserva Ducke, 26 km NE Manaus, 1 ♂ (MZSP 49199), 18.VI.1996, J.C.G. Hurtado leg. (MZSP).

FRENCH GUIANA (new country record)

– Amazone Nature Lodge, 4°33'N, 52°12'W, 980 ft., Montagne de Kaw, 1 ♂, 19-31.VIII.2019, Wappes & Morris leg. (FSCA).

***Inermestola boliviensis* sp. nov.**

(Fig. 27-31)

ZooBank: <http://zoobank.org/211F8C61-56D6-46BF-B368-CBB1E299E09E>

Holotype, ♂, BOLIVIA, Santa Cruz: 3.7 km SSE Buena Vista, Hotel Flora & Fauna, 430 m (blacklight trap, transition forest), 5-15.XI.2001, M.C. Thomas & B.K. Dozier leg. (FSCA).

Description of the holotype

Coloration. – Integument mostly black; anteclypeus dark yellowish brown anteriorly; labrum yellowish brown anteriorly; palpi reddish brown, except yellowish brown apex of maxillary palpomere IV and labial palpomere III; scape dark brown, slightly dark reddish brown on apex; pedicel dark reddish brown; antennomere III orangish brown on basal third, dark brown on remaining surface; antennomere IV yellowish on basal half, dark brown on apical half; antennomere V dark reddish brown basally, dark brown on remaining surface (missing antennomeres VI–XI on both antennae). Femora reddish brown apically, more distinctly dorsally. Posterior third of tarsomeres V and claws reddish brown. Ventrites 1–4 narrowly yellowish on apex; ventrite 5 reddish brown on sides of posterior third.

Head. – Frons coarsely, abundantly punctate; with short, decumbent, thick yellow setae not obscuring integument, and short, fine, decumbent white setae interspersed; with one long, erect brown seta on each side close to eyes. Vertex and area behind upper eye lobes coarsely, abundantly punctate; area between antennal tubercles and upper eye lobes with abundant, decumbent, thick yellow setae, and short, decumbent, fine white setae interspersed; central area of vertex between eyes and prothorax with longitudinal band of decumbent, thick yellow setae and remaining surface with fine, decumbent white setae in punctures; area behind upper eye lobes with dense yellow pubescence close to eye, and remaining surface with short, decumbent white setae in punctures. Area behind lower eye lobes coarsely, abundantly punctate; with abundant, decumbent, thick yellow setae not obscuring integument, and short, fine, decumbent white setae interspersed. Genae coarsely punctate except smooth glabrous apex; with decumbent, yellow and white setae not obscuring integument. Wide central area of postclypeus with yellow and white setae not obscuring integument, remaining surface with sparse white setae; with a few long, erect brownish setae. Sides of postclypeus glabrous. Labrum coplanar with anteclypeus at posterior 2/3, inclined at anterior third; posterior 2/3 with sparse yellowish-white pubescence, and long, erect brownish setae interspersed. Distance between upper eye lobes 0.30 times distance between outer margins of eyes; in frontal view, distance between lower eye lobes 0.51 times distance between outer margins of eyes. Antennae, from base of scape to apex of antennomere V, 0.8 times elytral length, reaching middle of elytra. Scape coarsely, somewhat rugose-punctate; with decumbent, thick yellow setae not obscuring integument, and decumbent, fine white setae interspersed. Pedicel and antennomeres III–V with short, decumbent white setae distinctly not obscuring integument.

Antennal formula based on length of antennomere III:

– Scape = 0.81. – Pedicel = 0.23. – IV = 1.19. – V = 0.69.

Thorax. – Prothorax transverse, with distinct posterior constriction. Sides of prothorax with sculpturing as on pronotum; with yellow setae as on pronotum, forming small isolated tufts throughout, and short white setae interspersed among them. Pronotum distinctly inclined on posterior quarter; coarsely, deeply, abundantly punctate; with short, thick yellow setae distinctly

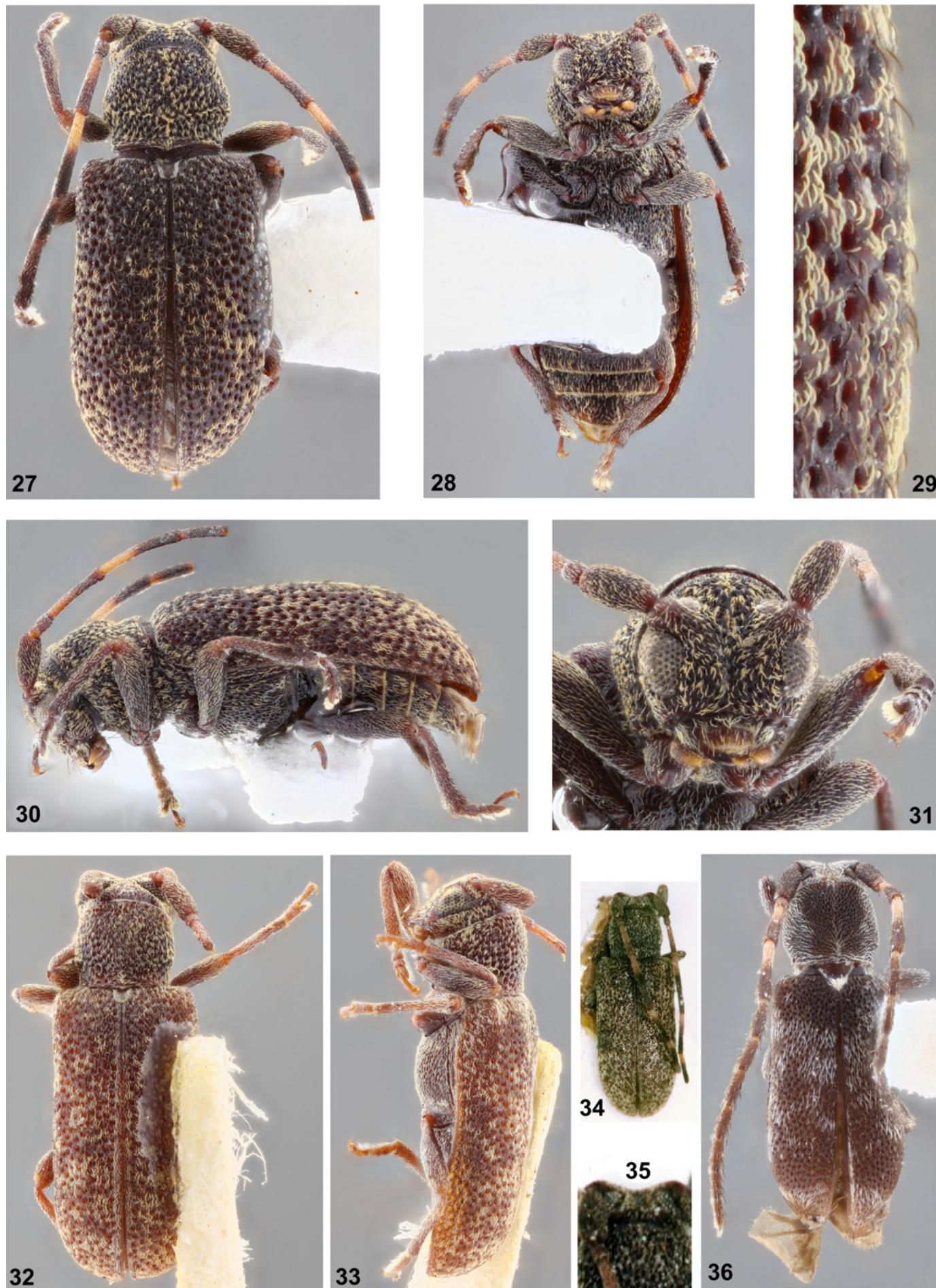


Fig. 27-36. *Inermestola* spp.

27-31. *Inermestola boliviensis* sp. nov., holotype ♂. 27. Dorsal habitus. 28. Ventral habitus. 29. Elytral erect setae. 30. Lateral habitus. 31. Head, frontal view.
 32-35. *Inermestola densepunctata* Breuning, 1942. 32. ♂, dorsal habitus. 33. ♂, lateral habitus. 34. Holotype, dorsal habitus (by Jesus Santiago Moure).
 35. Holotype, upper eye lobes (by Jesus Santiago Moure).
 36. *Inermestola chiapasensis* Galileo & Moysés, 2013, paratype ♂, dorsal habitus.

not obscuring integument, forming small isolated tufts, except on center of anterior and posterior third, where they are arranged in a band; with short, decumbent white setae interspersed on some areas. Prosternum with sculpturing and setae as on sides of prothorax. Prosternal process with moderately sparse white setae anteriorly and posteriorly, and yellow setae centrally; narrowest area 0.22 times procoxal width. Ventral surface of meso- and metathorax coarsely, abundantly punctate; mesoventrite with short, decumbent, somewhat sparse yellowish white setae except sides mostly with thick, decumbent yellow setae; mesanepisternum, mesepimeron, metanepisternum, and sides of metaventrite with short, thick, somewhat abundant yellow setae, and short, decumbent, fine white setae interspersed; remaining surface of metaventrite with short, decumbent, moderately sparse white setae, and short, decumbent yellow setae interspersed. Scutellum glabrous centrally, with dense grayish white pubescent band on margins.

Elytra. – Coarsely, abundantly, deeply punctate; with short, decumbent, thick yellow setae, denser from basal third, especially laterally and apically, and short, decumbent white setae interspersed on some areas, except basal area mostly with short white setae; with short, sub-erect brownish setae throughout.

Legs. – Femora and tibiae with short, decumbent, both yellow and white setae not obscuring integument, except posterior half of ventral surface of protibiae, about posterior third of dorsal surface of meso- and metatibiae with thick, bristly brown setae, and posterior third of ventral surface of meso- and metatibiae with bristly brownish setae; mesotibiae strongly sulcate dorsally. Metatarsomere I distinctly shorter than II–III together.

Abdomen. – Ventrites somewhat coarsely and abundantly punctate; sides with short, decumbent yellow setae not obscuring integument and whitish setae interspersed, and central area mostly with short, decumbent, sparse whitish setae. Apex of ventrite 5 rounded.

Dimensions (mm)

- Total length, 3.90)
- Prothoracic length, 0.85)
- Anterior prothoracic width, 0.90)
- Posterior prothoracic width, 0.95)
- Maximum prothoracic width, 1.05)
- Humeral width, 1.55)
- Elytral length, 2.70

Etymology. – The species name “*boliviensis*” refers to the country where the holotype was collected. Currently it is the only known species of the genus to occur in Bolivia.

Remarks. – *Inermestola boliviensis* sp. nov. resembles the species of *Estolomimus* Breuning, 1940. However, according to Breuning (1940b) and Martins & Galileo (1997), one of the conspicuous features of *Estolomimus* is the presence of a tubercle on the sides of the prothorax. This feature excludes the new species from *Estolomimus*.

Martins & Galileo (1998) separated *Inermestola* Breuning, 1942 and *Piimuna* Martins & Galileo, 1998 in his key as follows (translated):

“Distance between upper eye lobes equal to half width of a lobe; sides of the prothorax without central gibbosity; vertex concave,” leading to *Inermestola*; “Distance between upper eye lobes equal to three times the width of a lobe; sides of the prothorax with central gibbosity; vertex flat,” leading to *Piimuna*.

The new species has the body robust, as in *Piimuna gibbosa* Martins & Galileo, 1998 (the only known species of the genus), upper eye lobes closer to each other, as in *Inermestola densepunctata* Breuning, 1942, and the area between the upper ocular lobes is concave, as in *I. densepunctata*. These features put the new species between the two genera. However, based on the distance between the upper eye lobes and depressed area between them, we think it agrees better with *Inermestola*. Nevertheless, the information regarding the distance between the upper eye lobes in *I. densepunctata* is not correct, it is wider than the width of an upper eye lobe, as it is possible to see including on the holotype (Fig. 34–35).

Inermestola boliviensis sp. nov. differs from *I. densepunctata* (Fig. 32–33) as follows:

- (i) body shorter and stouter (Fig. 27).
- (ii) posterior quarter of the pronotum distinctly depressed (Fig. 30).
- (iii) lateral projection of the prothorax more distinct (Fig. 27).

In *I. densepunctata*:

- (i) the body is longer and slender (Fig. 32).
- (ii) the posterior quarter of the pronotum is slightly depressed (Fig. 33).
- (iii) the lateral projection of the prothorax is slightly distinct (Fig. 32).

***Inermestola chiapasensis* Galileo & Moysés, 2013**

(Fig. 36)

Inermestola chiapasensis Galileo & Moysés, 2013: 148.

Moura & Von Groll, 2017: 452 (paratype).

Monné, 2022: 697 (cat.).

Remarks. – According to Galileo & Moysés (2013) (translated): “In the key for identifying the South American genera of Desmiphorini with the sides of the prothorax without tubercle (Martins & Galileo, 1998), item 11 refers to the genera *Inermestola* and *Piimuna* Galileo & Martins, 1998: “antennomere III without short setae on the inner side, longer than the scape and as long as the IV” based on the photographic slide of the holotype of *I. densepunctata*, and a male from Chapada dos Guimarães, Mato Grosso. This character should be confirmed when it is possible to examine additional material.” However, antennomere III in *I. densepunctata*, as well as in the new species described here, has no erect setae on the ventral surface of antennomere III. In fact, erect setae are absent on all antennomeres of the two other species of *Inermestola*, while they are present in the antennomeres of *I. chiapasensis*. Furthermore, the distance between upper eye lobes is distinctly larger than in the other species of *Inermestola*. Therefore, it will be necessary to study *I. chiapasensis* further to correctly allocate the species, which is beyond the scope of this work.

Material examined. – MEXICO, Chiapas: Parque Nacional Sumidero, 1,000', ♂ paratype, 20.V-13.VI.1990, B. Gill, H. & A. Howden leg. (Malaise trap) (MZSP).

***Estola compacta* Breuning, 1940**

(Fig. 37–39)

Estola compacta Breuning, 1940a: 64.

Blackwelder, 1946: 600 (checklist).

Breuning, 1963: 508 (cat.); 1974b: 95.

Monné, 1994: 44 (cat.).

Monné & Giesbert, 1994: 218 (checklist).

Monné, 2005: 396 (cat.).

Monné & Hovore, 2006: 245 (checklist).

Monné, 2022: 665 (cat.).

Remarks. – *Estola compacta* was described and remained known only by the holotype (Fig. 39) from Brazil (Pará). It is similar to *Estola basiflava* Breuning, 1943 (Fig. 40), but differs especially by the pronotal pubescence not very dense (dense in *E. basiflava*) and asperate punctures on basal quarter of the elytra (punctures not asperate in *E. basiflava*).

Material examined.

ECUADOR (new country record)

– Napo: Napo-Galeras Road, km 3, 1 ♂ (Fig. 37), 1 ♀ (Fig. 38), 20.II.2004, F.T. Hovore leg. (CASC).

PERU (new country record)

– Junín: Satipo, 1 ♂, 1940, no collector indicated (MZSP).

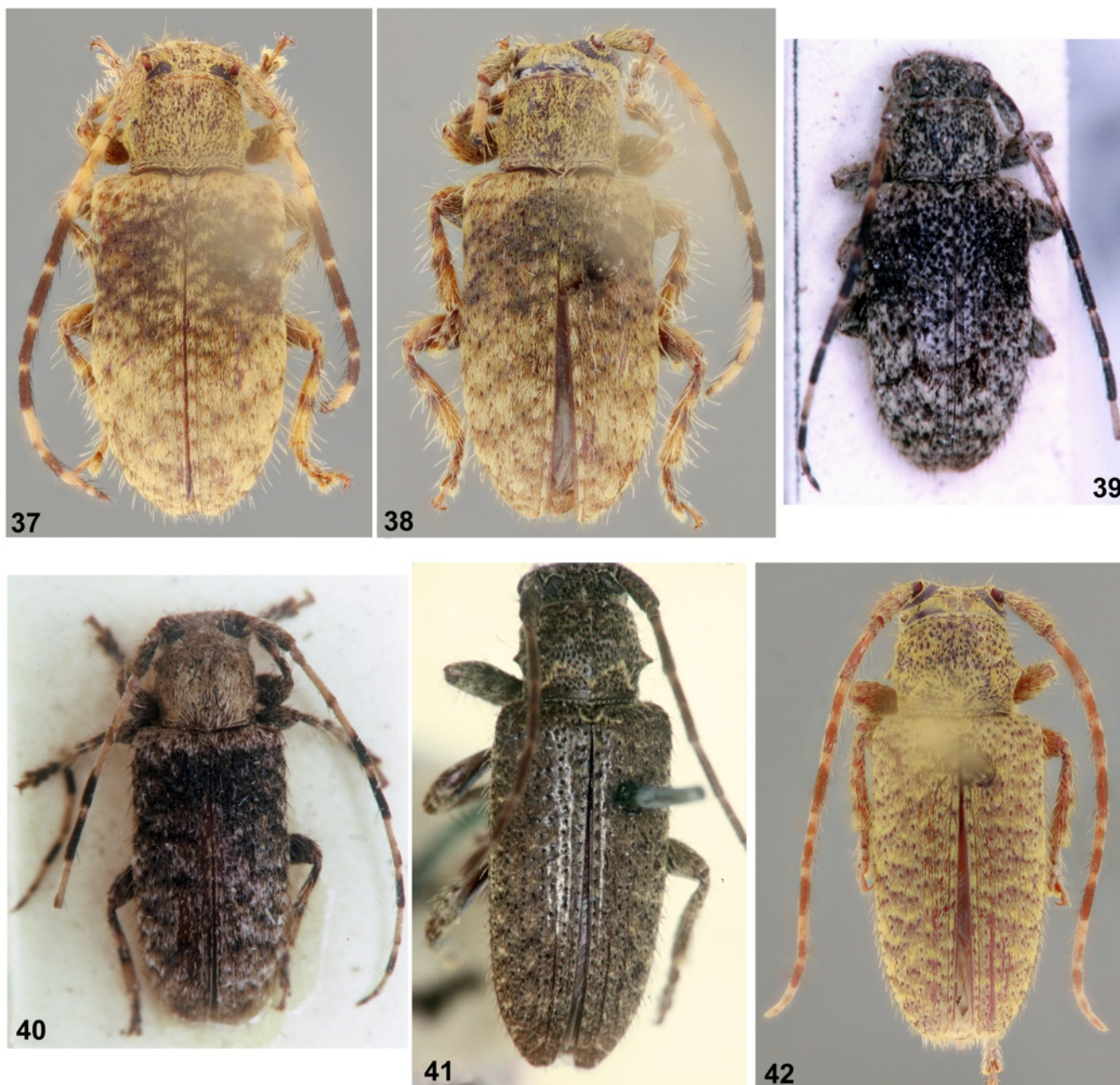


Fig. 37-42. *Estola* spp.

37-39. *Estola compacta* Breuning, 1940, dorsal habitus. 37. ♂, Ecuador. 38. ♀, Ecuador. 39. Holotype, by Jesus Santiago Moure.
 40. *Estola basiflava* Breuning, 1943.
 41-42. *Estola m-flava* Breuning, 1940, dorsal habitus. 41. Holotype, by Jesus Santiago Moure. 42. ♀, Trinidad and Tobago.

***Estola m-flava* Breuning, 1940**
 (Fig. 41-42)

Estola m-flava Breuning, 1940a: 64.
 Blackwelder, 1946: 601 (checklist).
 Breuning, 1963: 508 (cat.); 1974b: 82.
 Monné & Giesbert, 1994: 219 (checklist).
 Monné, 1994: 47 (cat.); Tavakilian *et al.*, 1997: 344 (hosts).
 Monné, 2002: 14 (cat.; hosts); 2005: 399 (cat.).
 Monné & Hovore, 2006: 245 (checklist).
 Morvan & Morati, 2006: 44 (distr.).
 Touroult *et al.*, 2010: 32.
 Martins *et al.*, 2011: 287 (distr.).
 Giuglaris, 2012: 64 (distr.).
 Morvan & Roguet, 2013: 26 (distr.).
 Monné, 2022: 670 (cat.).

Remarks. – Breuning describe *E. m-flava* based on a single specimen from French Guiana (Fig. 41). Breuning (1963) also

listed Brazil, without further details. Currently, *E. m-flava* is known from French Guiana and Brazil (Amazonas, Maranhão) (Monné 2022; Tavakilian & Chevillotte 2021). However, Breuning (1974b) listed also Santo Antonio da Barra, a city in the Brazilian state of Goiás.

Material examined.

TRINIDAD AND TOBAGO (new country record)

– Trinidad Island: Simla, Res. Sta. [Research Station], 1 ♀, 2-15.VI.1981, 1 ♂, Hanson & Clemons leg. (LGBC).

BRAZIL

Amapá (new state record):

– Rio Regina, Serra Lombard, 1 ♂ (MZSP 49205), 5.IX.1961, J. Bechyně leg. (MZSP).

Pará (new state record):

– Paragominas, 3 ♂ (MZSP 49200; MZSP 49203; MZSP 49204), 2.VI.1966, F.S. Pereira leg. (MZSP).
 – Serra Norte, Fofoca, 1 ♂ (MZSP 49202), 19-23.IX.1985, J. Dias leg. (MZSP).

– Santarém, Taperinha, 1 ♂ (MZSP 49206), 1920, Hagmann leg. (MZSP).

Maranhão:

– Bom Jardim, REBIO-Res. Biol. Gurupi, 1 ♀ (MZSP 49201), 5-15.VI. 2010, J.C. Silva, J.A. Silva, A.A. Santos & T.T.A. Silva leg. (MZSP).

Estola linearis sp. nov.

(Fig. 43-46)

ZooBank: <http://zoobank.org/DD7D9821-28AF-4C32-AD7B-BCD65A184B75>

Holotype, ♂, ECUADOR, *Orellana*: 16 km W Coca, 20.II.2004, F.T. Hovore leg. (CASC).

Description of the holotype

Coloration. – Integument mostly black; ventral mouthparts reddish brown, except yellowish brown apex of palpomeres; anteclypeus dark yellowish brown; basal fifth of antennomere VI orangish brown; basal 3/4 of antennomere VIII, basal fifth of antennomere IX, basal third of antennomere X, basal fifth and apex of antennomere XI orangish, anterior area of labrum yellowish brown; tarsomeres II dark brown; tarsomeres III–V and claws orangish brown; apex of ventrites 1–4 dark yellowish brown.

Head. – Frons moderately coarsely and abundantly punctate, except smooth area close to median groove; with both brownish and whitish pubescence not obscuring integument, except glabrous smooth area, and tuft of white pubescence on each side of central area, and fringe of white pubescence close to eyes; with long, erect, dark brown setae interspersed (some setae with whitish apex). Vertex with sculpturing as on frons; area between antennal tubercles with sparse whitish pubescence on central area, except glabrous smooth central area, both brownish and whitish pubescence not obscuring integument, and long, erect brown setae interspersed laterally; area between upper eye lobes with white pubescence close to eyes, denser toward antennal tubercles, both brownish and whitish pubescence not obscuring integument centrally, except glabrous smooth anterocentral area, tuft of pale yellow pubescence centrally, and long, erect brown setae interspersed (some with whitish apex); area close to prothorax with brownish pubescence, and sparse, short whitish setae interspersed. Area behind upper eye lobes moderately coarsely, abundantly punctate (punctures slightly finer than on vertex); with whitish pubescence partially obscuring integument. Area behind lower eye lobes coarsely, moderately sparsely punctate; with whitish pubescent band close to eye, distinctly widened and denser close to inferior margin, and abundant yellowish white pubescence not obscuring integument on remaining surface. Genae finely punctate close to eyes, smooth on remaining surface; with yellowish white pubescence close to eye (denser toward clypeus), glabrous on apex. Wide central area of postclypeus with yellowish white pubescence not obscuring integument, tuft of bristly whitish pubescence on center, and one long, erect dark brown seta on each side (apex of setae whitish); sides of postclypeus glabrous. Labrum coplanar with anteclypeus at posterior third, inclined at anterior third; finely, abundantly punctate; with sparse yellowish white pubescence and long, erect, both yellowish brown and yellowish white setae laterally; anterior margin with fringe of yellowish brown setae. Gulamentum smooth, glabrous, except narrow anterocentral area with grayish white pubescence. Distance between upper eye lobes 0.22 times distance between outer margins of eyes; in frontal view, distance between lower eye lobes 0.52 times distance between outer margins of eyes. Antennae 1.5 times elytral length, reaching elytral apex at posterior third of antennomere X. Scape finely, abundantly punctate; with sparse yellowish brown pubescence dorsally, partially brownish basally, moderately sparse whitish pubescence on sides, and dense whitish pubescence ventrally; with long, erect, dark brown setae interspersed dorsally (apex of erect setae whitish), and long, erect, sparse yellowish setae interspersed ventrally. Pedicel with brownish pubescence not obscuring integument, dorsally, whitish pubescence on remaining surface, and long, erect yellowish white setae interspersed ventrally. Antennomere III with brown pubescence not obscuring integument dorsally, abundant whitish pubescence on remaining surface, and long, erect yellowish brown setae interspersed ventrally (some of them dark brown basally). Antennomere IV with brown pubescence not obscuring integument dorsally, abundant whitish pubescence on basal quarter of sides and basal third of ventral surface,

brown pubescence with whitish pubescence interspersed on remaining surface, and long, erect dark brown setae interspersed ventrally. Antennomere V with brown pubescence not obscuring integument dorsally, whitish pubescence on base of sides and ventral surface, brown pubescence with moderately abundant whitish pubescence interspersed on remaining surface, and long, erect dark brown setae interspersed ventrally. Antennomeres VI and IX with basal ring of whitish pubescence, brown pubescence on remaining surface, and long, erect dark brown setae interspersed ventrally. Antennomere VII with narrow, sparse whitish pubescent ring basally, brown pubescence not obscuring integument on remaining surface, and long, erect dark brown setae interspersed ventrally. Antennomere VIII with dense white pubescence on basal 3/4, brownish pubescence not obscuring integument on posterior quarter, a few long, erect white setae interspersed ventrally on basal 3/4, and one long, erect dark brown seta on ventral surface of posterior quarter. Antennomere X with white pubescent ring on basal third, brown pubescence not obscuring integument on remaining surface, and a few long, erect dark brown setae interspersed ventrally. Antennomere XI with white pubescent ring basally and apically, and bristly brownish pubescence on remaining surface.

Antennal formula based on length of antennomere III:

– Scape = 1.40. – Pedicel = 0.33. – IV = 1.40. – V = 1.20. – VI = 1.13. – VII = 1.00. – VIII = 0.93. – IX = 0.87. – X = 0.73. – XI = 0.70.

Thorax. – Prothorax wider than long; sides with short, conical tubercle centrally. Sides of prothorax coarsely, abundantly punctate; lateral tubercles with dense pale yellow pubescence; remaining surface with abundant yellowish white pubescence not obscuring integument. Pronotum coarsely, abundantly punctate, except smooth anterior and posterior narrow area close to margins; wide central area with abundant brown pubescence not obscuring integument, and whitish pubescence interspersed (whitish pubescence denser centrally, anterior fifth, and center of area close to posterior margin); sides with wide whitish pubescent band, not reaching sides of prothorax anteriorly, and remaining surface of sides with pubescence as on central area; with long, erect, thick dark brown setae interspersed throughout. Prosternum coarsely, abundantly punctate; with bristly, abundant yellowish white pubescence partially obscuring integument, and long, erect yellowish brown setae interspersed. Prosternal process with sculpturing as on prosternum; with whitish pubescence centrally, longer on posterior area, and brown pubescence not obscuring integument on remaining surface; narrowest area 0.2 times procoxal width. Mesoventrite with sparse whitish pubescence centrally, dense, yellowish white laterally. Mesanepisternum and mesepimeron with dense yellowish white pubescence partially obscuring integument. Mesanepisternum coarsely, sparsely punctate; with moderately abundant whitish pubescence and long, suberect yellowish setae interspersed on posterior half. Metanepisternum and metaventrite with abundant grayish white pubescence partially obscuring integument, except sparse pubescence on center of metaventrite. Scutellum with abundant brown pubescence not obscuring integument.

Elytra. – Coarsely, sparsely punctate on basal third, punctures finer on remaining surface; with white pubescence forming two longitudinal bands on anterior 4/5, one wide, not distinctly dense dorsally close to suture (not reaching suture on basal quarter), another dense laterally; posterior fifth with white pubescence and brownish pubescent maculae interspersed; dorsal surface with longitudinal brown pubescent band between two white pubescent maculae on anterior 4/5, area close to suture on basal quarter, and area close to epipleura with brown pubescence and white pubescent maculae interspersed on anterior 4/5; with long, erect, thick dark brown setae interspersed throughout; apex rounded.

Legs. – Femora with abundant whitish pubescence not obscuring integument, slightly yellowish white apically, and long, erect, thick dark brown setae interspersed on posterior dorsal half. Tibiae with bristly white pubescence not obscuring integument, except yellowish brown bristly pubescence on posterior third of ventral surface, and pale yellow pubescence on dorsal sulcus of mesotibiae; protibiae with long, erect yellowish white setae interspersed dorsally and laterally (some of erect setae dark brown basally); meso- and metatibiae with long, erect dark brown setae interspersed, distinctly more abundant on dorsal posterior half. Metatarsomere I shorter than II–III together.



Fig. 43-47. *Estola* spp.

43-46. *Estola linearis* sp. nov., holotype ♂. 43. Dorsal habitus. 44. Ventral habitus. 45. Lateral habitus. 46. Head, frontal view. 47. *Estola operosa* Martins & Galileo, 2007, holotype ♂, dorsal habitus.

Abdomen. – Ventrites 1–4 with grayish white pubescence, slightly longer, forming longitudinal band centrally (less distinctly on 1), sparser on sides of central band, two dense grayish white pubescent bands laterally, with brownish pubescent band between them; ventrite 5 with abundant grayish white pubescence, slightly sparser on anterocentral half; with long, erect sparse, mostly yellowish brown setae on 1–4, and long, erect dark brown setae on 5.

Dimensions (mm)

- Total length, 6.65
- Prothoracic length, 1.20
- Anterior prothoracic width, 1.35
- Posterior prothoracic width, 1.40
- Maximum prothoracic width, 1.60
- Humeral width, 1.95
- Elytral length, 4.65

Etymology. – The species name “*linearis*” (Latin) refers to the longitudinal lines of light colored pubescence on the elytra.

Remarks. – *Estola linearis* sp. nov. is similar to *E. operosa* Martins & Galileo, 2007 (Fig. 47), but differs as follows:

- (i) antennomere III entirely dark.
- (ii) antennomere IV as long as scape, entirely dark.
- (iii) antennomere V entirely dark.
- (iv) antennomere X light only basally.
- (v) prothorax distinctly wider than long.
- (vi) lateral tubercles of prothorax with dense pale yellow pubescence.
- (vii) scutellum with brown pubescence.
- (viii) elytra with longitudinal brown pubescent band dorsally, slightly narrowed posteriorly.

In *E. operosa* :

- (i) the posterior third of antennomere III is dark reddish brown.
- (ii) antennomere IV is distinctly longer than scape, pale yellow on basal 2/3.
- (iii) antennomere V pale yellow on basal third, reddish brown on central area.
- (iv) antennomere X almost entirely pale yellow.
- (v) prothorax is slightly wider than long.
- (vi) lateral tubercles of the prothorax without yellow pubescence.
- (vii) scutellum with dense whitish pubescence.
- (viii) elytra without longitudinal brown pubescent band dorsally, and distinctly narrowed posteriorly.

The new species differs from *E. albovaria* Breuning, 1940 and *E. nigrescens* Breuning, 1943 (see photographs on Bezark 2022) by the elytral pubescence forming longitudinal bands, notably on the sides (not forming bands in *E. albovaria* and *E. nigrescens*); it also differs from *E. albovaria* by the antennomere III shorter than the scape (as long as scape in *E. albovaria*), and the pronotum without a narrow and very dense pubescent band on each side of the base (present in *E. albovaria*).

***Eupogonius fuscovittatus* Breuning, 1974**

(Fig. 48-52)

Eupogonius fuscovittatus Breuning, 1974a: 26; 1974b: 154.

Chemsak *et al.*, 1992: 119 (cat.).

Monné, 1994: 30 (cat.).

Monné & Giesbert, 1994: 221 (checklist).

Monné, 2005: 409 (cat.).

Hovore, 2006: 376 (distr.).

Monné, 2022: 687 (cat.).

Remarks. – Breuning, (1974a) described *E. fuscovittatus* based on a single specimen (Fig. 52) from Guatemala (Zapote). Subsequently, the species has only been mentioned in catalogs and checklists. We take the opportunity to illustrate the species in detail (Fig. 48-51). This species is easily recognized among its congeners by the denuded elongate longitudinal vittae on the elytra placed near the suture, as well as the group of white setae at the base of only the fourth antennal segment which contrast with the remaining setae throughout the antennae.

Material examined. – GUATEMALA, *Suchitepequez* (new department record): 11 km N Patulul, 700-1000 m, 14°31'N, 91°08'W, 1 ♂, 10-13.VI.2012, E. Fuller leg. (LGBC).

***Estoloides elongata* sp. nov.**

(Fig. 53-57)

ZooBank: <http://zoobank.org/22BCC4D1-6DA2-4A54-AD17-7659FC6021E5>

Holotype, ♂, PANAMA, *Panama*: El Llano, 13.V.1991, F.T. Hovore leg. (CASC).

Paratypes

– 1 ♀, same data as holotype (LGBC).

– 1 ♂, PANAMA, *Panama*: Cerro Jefé, 2200', 21-27.V.1996, Wappes, Huether & Morris leg. (MZSP).

Description of the holotype (Fig. 53-56)

Coloration. – Integument mostly brown dorsally, mostly dark brown ventrally; ventral mouthparts mostly dark reddish brown, except mentum dark brown, and apex of palpomeres yellowish brown; anteclypeus yellowish brown; scape, pedicel, and antennomere III reddish brown, darker on apex of antennomere III; remaining antennomeres dark orangish brown. Femora and tibiae reddish brown, except dark reddish brown base of tibiae; tarsi mostly dark brown.

Head. – Frons coarsely, abundantly punctate; with dense orangish brown pubescence partially obscuring integument, not obscuring

punctures; with moderately long, erect yellowish white setae interspersed, and long, erect yellowish white setae close to eyes. Area between antennal tubercles and posterior margin of upper eye lobes coarsely, sparsely punctate; with dense orangish brown pubescence obscuring integument on some areas; remaining surface of vertex smooth, with dense orangish brown pubescence partially obscuring integument; with a few long, erect yellowish white setae interspersed between eye lobes. Area behind upper eye lobes coarsely, sparsely punctate (punctures finer and sparser than on anterior area of vertex); with dense orangish brown pubescence almost obscuring integument, and a few long, erect whitish setae close to eye. Area behind lower eye lobes tumid, coarsely, sparsely punctate close to eye, finely, shallowly, abundantly punctate close to prothorax; with dense pale yellow pubescence on tumid area, almost glabrous close to prothorax, and with long, erect yellowish white setae close to eye. Genae with abundant yellowish pubescence not obscuring integument, sparser toward apex, and long, erect yellowish white setae interspersed. Wide central area of postclypeus coarsely, sparsely punctate; with orangish brown pubescence not obscuring integument close to frons, and fringe of sparse yellowish white setae close to anteclypeus; with long, erect yellowish white setae interspersed, more abundant close to anteclypeus. Sides of postclypeus smooth, glabrous. Labrum coplanar with anteclypeus at posterior third, inclined at anterior 2/3; finely, abundantly punctate; with yellowish white pubescence distinctly not obscuring integument on coplanar area; with long yellowish-brown setae directed forward (apex of nearly all setae whitish apically); anterior margin with fringe of short yellowish brown setae. Gula smooth, glabrous, except narrow anterior area coarsely, sparsely punctate, with yellowish pubescence, and long, erect yellowish white seta on each puncture. Distance between upper eye lobes 0.3 times distance between outer margins of eyes; in frontal view, distance between lower eye lobes 0.5 times distance between outer margins of eyes. Antennae 1.85 times elytral length, reaching elytral apex at posterior sixth of antennomere VIII. Scape with abundant yellowish brown pubescence dorsally and laterally not obscuring integument, and dense whitish pubescence ventrally; with a few long, erect yellowish brown setae interspersed, distinctly longer and more abundant ventrally. Pedicel with pubescence as on scape and long, erect yellowish white setae ventrally (erect setae brownish basally). Antennomeres III–XI with basal white pubescent ring, remaining surface with yellowish pubescence not obscuring integument, and short, erect, sparse yellowish setae interspersed throughout; antennomeres III–VII with long, erect yellowish white setae ventrally, distinctly more abundant on III, gradually shorter and sparser toward VII; apex of antennomere XI strongly arched inward and almost forming the 12th segment.

Antennal formula based on length of antennomere III:

– Scape = 0.80. – Pedicel = 0.15. – IV = 1.10. – V = 0.85. – VI = 0.80. – VII = 0.78. – VIII = 0.73. – IX = 0.71. – X = 0.65. – XI = 0.68.

Thorax. – Prothorax distinctly wider than long; sides with large conical tubercle on middle. Pronotum coarsely, moderately abundantly punctate; with dense orangish brown pubescence partially obscuring integument, not obscuring punctures, except yellowish white pubescence close to posterior margin, and grayish green pubescence on lateral tubercles of prothorax; with long, erect, sparse yellowish white setae interspersed. Sides of prothorax coarsely, sparsely punctate; with grayish green pubescence partially obscuring integument on wide superior area, except yellowish white pubescence close to posterior margin, and yellowish white pubescence partially obscuring integument close to prosternum. Prosternum moderately coarsely, sparsely punctate on posterior 2/3, impunctate on anterior third; with abundant grayish yellow pubescence not obscuring integument, slightly yellowish centrally, and long, erect yellowish setae interspersed on posterior 2/3. Prosternal process with pubescence as on prosternum; narrowest area 0.43 times procoxal width. Ventral surface of meso- and metathorax with abundant grayish yellow pubescence partially obscuring integument (slightly yellowish on mesepimeron and metanepisternum, and more grayish white on remaining surface depending on light intensity); metaventricle with long, erect, sparse yellowish white setae interspersed. Scutellum glabrous on anterocentral area, with dense yellow pubescence on remaining surface.



Fig. 48–52. *Eupogonius fuscovittatus* Breuning, 1974.

48–51. ♂, Guatemala, Suchitepequez. 48. Dorsal habitus. 49. Ventral habitus. 50. Lateral habitus. 51. Head, frontal view. 52. Holotype, dorsal habitus, by John Chemsak.

Elytra. – Coarsely, abundantly punctate on basal third, punctures gradually finer, sparser toward apex; with dense orangish brown pubescence obscuring integument, not obscuring punctures; each puncture with short, subdecumbent dark brown seta; sides with long, erect yellowish white setae interspersed; apex rounded.

Legs. – Femora with abundant grayish white pubescence not obscuring integument, and long, erect yellowish white setae interspersed dorsally. Protibiae with abundant grayish white pubescence not obscuring integument, except posterior half of ventral surface with dense, bristly yellowish brown pubescence, and long, erect setae dorsally and laterally (setae dark brown basally, gradually yellowish toward apex). Mesotibiae with abundant grayish white pubescence not obscuring integument (more yellowish depending on light source and intensity), except posterior half of dorsal surface with short, thick, erect, abundant yellowish brown setae (darker depending on light source), and posterior third of ventral surface with short, suberect yellowish brown setae; with long, erect yellowish white setae interspersed laterally (longer and more abundant on outer surface). Metatibiae with abundant yellowish white pubescence not obscuring integument; posterior half with short, erect, thick, abundant yellowish brown setae interspersed; with long, erect yellowish white setae interspersed throughout. Metatarsomere I about as long as II–III together.

Abdomen. – Ventrites with abundant grayish white pubescence partially obscuring integument, except glabrous apex of ventrites 1–4, and fringe of yellowish white setae close to glabrous area of ventrites 1–4; with long, erect, sparse yellowish white setae interspersed; apex of ventrite 5, widely emarginate centrally.

Female (Fig. 57). – Differs from male only by the shorter antennae, 1.3 times elytral length, slightly surpassing elytral apex, and apex of antennomere XI not arched inward.

Dimensions (mm) (holotype ♂ / paratype ♂ / paratype ♀).

- Total length, 16.00/15.80/17.55
- Prothoracic length, 2.95/2.80/3.05
- Anterior prothoracic width, 2.95/2.70/3.05
- Posterior prothoracic width, 3.45/3.05/3.60
- Maximum prothoracic width, 4.45/4.20/4.50
- Humeral width, 5.10/4.70/5.40
- Elytral length, 11.65/11.20/13.10

Etymology. – The specific epithet “*elongata*” (Latin) refers to the overall shape of the species as compared to the similar species discussed below.

Remarks. – *Estoloides elongata* sp. nov. is similar to *E. perforata* (Bates, 1872) (Fig. 58-59), but differs as follows:

- (i) elytra proportionally longer, about 4.0 times prothoracic length in ♂, about 4.3 times in ♀.
- (ii) antennae distinctly shorter, reaching elytral apex near apex of antennomere VIII in ♂, and slightly surpassing elytral apex in ♀.
- (iii) antennomere III shorter than prothorax in ♂.
- (iv) apex of antennomere XI distinctly arched inward in ♂.
- (v) basal white pubescent ring on flagellomeres well-delimited.

In *E. perforata* :

- (i) elytra proportionally shorter, about 3.3 times prothoracic length in ♂, and 4.0 in ♀.
- (ii) antennae distinctly longer, surpassing elytral apex about apex of antennomere VI in ♂, distinctly surpassing elytral apex in ♀.
- (iii) apex of the antennomere XI not arched inward in ♂.
- (iv) antennomere III longer than prothorax in ♂.
- (v) basal whitish pubescent ring not well-delimited, especially toward distal segments.

According to Santos-Silva *et al.* (2018) on *E. perforata*: “A feature that best helps recognition of *E. perforata* is the grayish pubescence on ventral side of the body, as well as on the sides of humerus, basal third of the epipleura (distinctly contrasting with remaining elytral pubescence), and sides of prothorax [...] it is likely that *E. perforata* is a complex of very similar species. However, if so, no character pointed out by Bates (1872, 1885), or Breuning (1940a, 1974b), nor found by us, can consistently separate them.” Among the features pointed out by Breuning (1940a, 1974b) separating *E. longicornis* Breuning, 1940 (Fig. 60-61) from *E. perforata* (Bates, 1872) only the color of the pubescence on ventral side of the body can, eventually, separate these two species. According to Breuning (1940a, 1974b) the pubescence is brownish yellow in *E. longicornis*. If the description of the ventral pubescence in *E. longicornis* by Breuning (1940a, 1974b) is accurate, then the new species differs from *E. longicornis* by the same features pointed out separating it from *E. perforata*, and also due to the color of the ventral pubescence.

***Estolomimus ochreoapicalis* (Breuning, 1973) comb. nov.**

(Fig. 62-65)

Laraesima ochreoapicalis Breuning, 1973: 664.

Monné, 1980b: 232.

Breuning, 1980: 15; Monné, 1994: 20 (cat.).

Monné & Giesbert, 1994: 213 (checklist).

Monné, 2005: 367 (cat.).

Martins & Galileo, 2013: 321 (distr.).

Nascimento *et al.*, 2017: 91 (distr.).

Monné, 2022: 621 (cat.).

Remarks. – Breuning (1973) described *Laraesima ochreoapicalis* based on three specimens from Brazil (Pernambuco). *Laraesima* Thomson, 1868 is a genus of Comptosomatini Thomson, 1857. However, the tarsal claws in Comptosomatini are divergent, while they are divaricate in *L. ochreoapicalis*. In fact, *L. ochreoapicalis* belongs in Desmiphorini in the genus *Estolomimus* Breuning, 1940.

Monné (1980b) redescribed *L. ochreoapicalis* based on four males and four females from Brazil (Pernambuco). However, we do not know if the specimens were misidentified or if the tarsal claws were examined. We have the same question regarding the female from Brazil (Paraíba) examined by Martins & Galileo (2013) and Nascimento *et al.* (2014). Currently, the species is reported only from Brazil (Pernambuco and Paraíba) (Monné 2022; Tavakilian & Chevillotte 2021).

HEMILOPHINI Thomson, 1868

***Eranina stephanus* (Aurivillius, 1900) comb. nov.**

(Fig. 66-67)

Amphionycha stephanus Aurivillius, 1900: 420.

Adesmus stephanus

Aurivillius, 1923: 591 (cat.).

Blackwelder, 1946: 624 (checklist).

Gilmour, 1965: 637 (cat.).

Monné, 1995: 28 (cat.).

Monné & Giesbert, 1994: 281 (checklist).

Galileo & Martins, 1999: 91.

Monné, 2005: 447 (cat.).

Martins & Galileo, 2014: 60.

Monné, 2022: 752 (cat.).

Remarks. – Aurivillius (1923) synonymized *Amphionycha* Dejean, 1835 with *Adesmus* Lepeletier and Audinet-Serville, 1825. Consequently, *Amphionycha stephanus* was transferred to *Adesmus*. Galileo & Martins (1999) did not examine specimens of *A. stephanus* but provided a drawing based on the photographic slides taken by Jesus Santiago Moure. However, it is practically impossible to recognize the species based on that drawing. We provide the original photographs taken by Jesus Santiago Moure (Fig. 66-67), which allow concluding that the species belongs to *Eranina* Monné, 2005. The species remains known only by the holotype from Venezuela.

***Eranina monnei* sp. nov.**

(Fig. 68-71)

ZooBank: <http://zoobank.org/2C63815C-639F-4887-9489-B6B06A7C3DA9>

Holotype, ♀, PANAMA, Coclé: El Valle, 640 m, 08°36.868'N, 80°06.779'W, 26-28.VI.2011, E. Fuller leg. (CASC, formerly LGBC).

Description of the holotype

Coloration. – Head capsule mostly dark reddish brown, except one wide, oblique band on each side of frons, narrow area close to lower eye lobes on frons, antennal tubercles, and wide longitudinal band involving sides of vertex and area behind upper eye lobes black; scape mostly black dorsally and mostly dark orangish brown laterally and ventrally; pedicel black; antennomere III orangish brown basally, dark brown toward posterior third on dorsal surface, dark reddish brown with irregular dark brown areas interspersed toward posterior third of sides and ventral surface, yellowish brown on posterior third; antennomeres IV–VI yellowish brown (lighter toward VI; missing antennomeres VII–XI). Pronotum black laterally, with wide, longitudinal dark reddish brown band centrally, from base to apex; sides of prothorax black close to pronotum, dark reddish brown toward prosternum; prosternum reddish brown; ventral surface of mesothorax reddish brown, except large black central macula on mesoventrite; metanepisternum reddish brown; metaventrite black, except one wide, oblique band on each side of middle and narrow area close to metanepisternum orangish brown. Elytra black, except longitudinal reddish brown band close to suture, and orangish brown apex. Coxae and femora orangish brown; tibiae black, except most of ventral surface of protibiae and macula on ventral surface of basal third of mesotibia reddish brown; tarsomeres I–II mostly dark brown; tarsomeres III–V mostly yellowish brown. Ventriles orangish brown with irregular reddish brown areas interspersed.

Head. – Frons moderately finely, abundantly punctate; with abundant whitish pubescence not obscuring integument, and long, erect yellowish white setae interspersed. Vertex moderately finely, abundantly punctate, except smooth area close to prothorax; with abundant yellowish white pubescence obscuring integument, except glabrous area close to prothorax and sparse, brownish pubescence



Fig. 53-61. *Estoloides* spp.

53-56. *Estoloides elongata* sp. nov., holotype ♂. **53.** Dorsal habitus. **54.** Ventral habitus. **55.** Lateral habitus. **56.** Head, frontal view.

57. *Estoloides elongata* sp. nov., paratype ♀, dorsal habitus.

58-59. *Estoloides perforata* (Bates, 1872), ♂, Costa Rica, Turrialba. **58.** Dorsal habitus. **59.** Lateral habitus.

60-61. *Estoloides longicornis* Breuning, 1940, holotype ♂. **60.** Oblique view, by Jesus Santiago Moure. **61.** Dorsal habitus, by Henry Hespenseide.

close to upper eye lobes; with long, erect yellowish white setae interspersed on wide central area. Area behind upper eye lobes with abundant brownish pubescence not obscuring integument. Area behind lower eye lobes with yellowish white pubescence close to upper eye lobes, gradually whitish, sparser toward ventral surface except glabrous area close to prothorax. Genae finely, sparsely punctate, except smooth apex; with yellowish white pubescence not obscuring integument, slightly more abundant toward clypeus, except glabrous apex; with a few long, erect yellowish white setae interspersed. Wide central area of postclypeus with sparse whitish pubescence except glabrous central region; with a few moderately long yellowish setae interspersed, and one long, erect yellowish seta on each side. Sides of postclypeus glabrous. Labrum coplanar with anteclypeus at posterior 2/3, inclined at anterior third; coarsely punctate on posterior 2/3, smooth on anterior third; with long, erect yellowish setae laterally. Gula mentum smooth, glabrous. Distance between upper eye lobes 0.27 times distance between outer margins of eyes; in frontal view, distance between lower eye lobes 0.59 times distance between outer margins of eyes. Antennae 1.35 times elytral length (from base of scape to apex of antennomere VI), reaching elytral apex near apex of antennomere VI. Scape with abundant, bristly dark yellowish brown pubescence dorsally, sparse, yellowish white laterally and ventrally; with long, erect yellowish brown setae interspersed, distinctly longer and more abundant ventrally. Pedicel with sparse yellowish white pubescence, a few moderately long, erect setae of same color interspersed dorsally, and a few long, erect yellowish brown setae interspersed ventrally. Antennomere III with bristly yellowish white pubescence on anterior and posterior light areas, sparse brownish pubescence dorsally on dark central area, absent on remaining dark central area; dorsal surface with short, erect, abundant brownish setae dorsally on dark central area, and long, erect brownish setae ventrally on anterior third. Antennomere IV with abundant, bristly whitish pubescence not obscuring integument, and a few long, erect setae of same color interspersed ventrally. Antennomeres V–VI with abundant, bristly whitish pubescence not obscuring integument.

Antennal formula based on length of antennomere III:

– Scape = 0.52. – Pedicel = 0.10. – IV = 0.73. – V = 0.48. – VI = 0.37.

Thorax. – Prothorax distinctly transverse; sides slightly divergent from anterolateral angles to posterior third, then subparallel-sided toward posterolateral angles. Pronotum coarsely, moderately abundantly punctate; with longitudinal yellowish white pubescent band centrally, pubescence sparser on anterior fifth; remaining surface with brown pubescence not obscuring integument; with long, erect, sparse yellowish setae interspersed throughout. Sides of prothorax

with sculpturing as on pronotum; with abundant yellowish white pubescence not obscuring integument, and long, erect setae of same color interspersed. Prosternum with sparse yellowish white pubescence. Narrowest area of prosternal process 0.2 times procoxal width. Ventral surface of meso- and metathorax with yellowish white pubescence not obscuring integument, sparser centrally; metaventrite with long, erect, sparse yellowish white setae interspersed. Scutellum with abundant yellowish white pubescence.

Elytra. – Coarsely, abundantly punctate on basal half, punctures gradually finer toward apex; sutural area with dense yellowish white pubescent band from scutellum to posterior third, slightly sparser from this area to apical orangish brown region (forming longitudinal pubescent band from area between antennal tubercles to apical orangish brown region of elytra); apical orangish brown region with abundant yellowish white pubescence partially obscuring integument; remaining anterior 2/3 with abundant yellowish white pubescence not obscuring integument; area between beginning of posterior third and apical orangish brown region with abundant brown pubescence not obscuring integument; with long, erect yellowish setae interspersed throughout, more brownish on area with brown pubescence; apex rounded.

Legs. – Femora with yellowish white pubescence not obscuring integument, and long, erect setae of same color interspersed, more abundant ventrally close to base; tibiae with yellowish white pubescence basally, bristly, yellowish brown on remaining surface, especially on meso- and metatibiae; with long, erect yellowish setae interspersed (slightly darker on meso- and metatibiae).

Abdomen. – Ventrites with yellowish white pubescence not obscuring integument, denser centrally on ventrites 1–4, and long, erect setae of same color interspersed; apex of ventrite 5 truncate.

Dimensions (mm).

- Total length, 6.65
- Prothoracic length, 1.00
- Anterior prothoracic width, 1.30
- Posterior prothoracic width, 1.45
- Maximum prothoracic width, 1.50
- Humeral width, 2.00
- Elytral length, 4.85

Etymology. – The species epithet is in honor of Miguel A. Monné, one of the greatest specialists on the taxonomy of New World Cerambycidae and author of the genus *Eranina*.



Fig. 62-65. *Estolomimus ochreoapicalis* (Breuning, 1973), holotype.

62. Tarsal claws, by Kimberly García. 63. Labels. 64. Dorsal habitus, by Jesus Santiago Moure. 65. Lateral habitus, by Kimberly García.



66



67



68



69



70



71

Fig. 66-71. *Eranina* spp.

66-67. *Eranina stephanus* (Aurivillius, 1900), holotype, by Jesus Santiago Moure. 66. Dorsal habitus. 67. Lateral habitus.

68-71. *Eranina monnei* sp. nov., holotype ♀. 68. Dorsal habitus. 69. Ventral habitus. 70. Lateral habitus. 71. Head, frontal view.

Remarks. – *Eranina monnei* sp. nov. is similar to *E. stephanus* (Fig. 66-67) by the presence of the longitudinal pubescent band from the vertex to posterior area of the elytra.

However, it differs as follows :

- (i) elytra without oblique pubescent band on the middle of the dorsal surface.
- (ii) elytral apex uniformly rounded.
- (iii) apical area of the elytra orangish brown.
- (iv) frons and sides of thorax without dense white pubescence.
- (v) tibiae mostly black.

In *E. stephanus* :

- (i) elytra with an oblique pubescent band on the middle of the dorsal surface, without light apical area.
- (ii) elytral apex truncate, with outer angle projected.
- (iii) frons and sides of thorax with dense white pubescence.
- (iv) tibiae orangish.

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References

- Aurivillius C., 1900. – Verzeichniss der von Dr. F. Meinert im Jahre 1891 in Venezuela gesammelten Cerambyciden. *Öfversigt Svenska Vetenskaps-Akademiens Förhandlingar*, 57: 409-421.
- Aurivillius C., 1923. – Cerambycidae: Lamiinae II, pp. 323-704. In: Junk, W. & Schenkling, S. (Eds.), *Coleopterorum Catalogus*, pars 74, W. Junk, Berlin, 381 pp.
- Bates H.W., 1872. – On the longicorn Coleoptera of Chontales, Nicaragua. *The Transactions of the Entomological Society of London*, 1872: 163-238.
- Bates H.W., 1881. – Longicornia, pp. 153–224. In: Godman, F.D. & Salvin, O. (eds.). *Biologia Centrali-Americana, Insecta, Coleoptera*. Vol. 5. Taylor and Francis, London, xii + 525 pp.
- Bates H.W., 1885. – Supplement to Longicornia, pp. 249-436. In: Godman, F.D. & Salvin, O. (eds.). *Biologia Centrali-Americana, Insecta, Coleoptera*. Vol. 5. Taylor and Francis, London, xii + 525 pp.
- Bezark L.G., 2021. – Checklist of the Oxypeltidae, Vesperidae, Disteniidae and Cerambycidae (Coleoptera) of the Western Hemisphere. 2021 Edition (updated through 31 December 2020). Available from: <http://bezbycids.com/byciddb/wdefault.asp?w=n/> (Last accessed 24 February 2022.)
- Bezark L.G., 2022. – A photographic Catalog of the Cerambycidae of the World. New World Cerambycidae Catalog. Available from <http://bezbycids.com/byciddb/wdefault.asp?w=n/> (Last accessed 24 February 2022.)
- Blackwelder R.E., 1946. – Checklist of the coleopterous insects of Mexico, Central America, the West Indies and South America. Part 4. *Bulletin of the United States National Museum*, 185: 551-763.
- Breuning S., 1940a. – Novae species Cerambycidae. VIII. *Folia Zoologica et Hydrobiologica*, 10: 37-85.
- Breuning S., 1940b. – Novae species Cerambycidae. IX. *Folia Zoologica et Hydrobiologica*, 10: 115-214.
- Breuning S., 1961. – *Catalogue des Lamiaires du Monde* (Col., Cerambycidae). *Verlag des Museum G. Frey, Tutzing bei München*, 5: 287-382.
- Breuning S., 1963. – *Catalogue des Lamiaires du Monde* (Col., Cerambycidae). *Verlag des Museum G. Frey, Tutzing bei München*, 7: 463-555.
- Breuning S., 1973. – Lamiaires nouveaux ou peu connus du Muséum de Paris (Col., Cerambycidae). *Annales de la Société Entomologique de France* (n.s.), 9(3): 647-665.
- Breuning S., 1974a. – Neue Arten und Gattungen von Lamiinen (Coleoptera, Cerambycidae). *Mitteilungen aus dem Zoologischen Museum in Berlin*, 50(1): 149-165.
- Breuning S., 1974b. – Révision des Rhodopinini américains. *Studia Entomologica*, 17(1-4): 1-210.
- Breuning S., 1980. – Révision de la tribu des Compsosomatini (Coleoptera, Cerambycidae). *Sciences Nat*, Venette, 32 pp.
- Chemsak J.A. & Linsley E.G., 1970. – Additional designations of lectotypes of neotropical Cerambycidae in the collections of the British Museum (Natural History) (Coleoptera). *Journal of the Kansas Entomological Society*, 43(4): 404-417.
- Chemsak J.A., Linsley E.G. & Noguera F.A., 1992. – Listados faunísticos de México. II. Los Cerambycidae y Disteniidae de Norteamérica, Centroamérica y las Indias Occidentales (Coleoptera). Universidad Nacional Autónoma, Mexico City, 204 pp.
- Galileo M.H.M. & Martins U.R., 1991. – Revisão da tribo Calliini (Coleoptera, Cerambycidae, Lamiinae). *Giornale Italiano di Entomologia*, 5: 243-262.
- Galileo M.H.M. & Martins U.R., 1998. – Notas e descrições em Lamiinae (Coleoptera, Cerambycidae) com garras tarsais apendiculadas ou bifidas e revisão do 1º grupo de espécies do gênero *Malacoscyllus* Thomson, 1868. *Revista Brasileira de Entomologia*, 41(2-4): 249-256.
- Galileo M.H.M. & Martins U.R., 1999. – O gênero *Adesmus* (Coleoptera, Cerambycidae, Lamiinae, Hemilophini). *Iheringia* (Zoologia), 86: 77-116.
- Galileo M.H.M. & Martins U.R., 2002. – Espécies novas e chave para as espécies de *Callia* (Coleoptera, Cerambycidae). *Iheringia* (Zoologia), 92: 41-52.
- Galileo M.H.M. & Moysés E., 2013. – *Blabicentrus* Bates, 1866 e *Inermestola* Breuning, 1942 (Cerambycidae, Lamiinae, Desmiphorini): espécies novas da região neotropical. *Papéis Avulsos de Zoologia*, 53(11): 145-149.
- Gilmour E.F., 1965. – *Catalogue des Lamiaires du Monde* (Col., Cerambycidae). *Museum G. Frey, Tutzing bei München*, 8: 559-655.
- Giuglaris J.L., 2012. – Les longicornes en zone littorale de Guyane: échantillonnage de la zone agricole Wayabo de Matiti (Coleoptera, Cerambycidae). *Bulletin de liaison de l'ACOREP*, 6(supplement): 53-65.
- Hovore F.T., 2006. – The Cerambycidae (Coleoptera) of Guatemala, pp. 363–378. In: Cano, E. (Ed.), *Biodiversidad de Guatemala*. Vol. 1. Universidad del Valle de Guatemala, Guatemala City, i-vi + 674 pp.



Fig. 72. Belize, Las Cuevas, Cayo District, near the Guatemalan border.

- Martins U.R. & Galileo M.H.M., 1997. – Revisão dos gêneros *Pseudestola* Breuning, *Estolomimus* Breuning e *Euestola* Breuning (Coleoptera, Cerambycidae, Lamiinae, Desmiphorini). *Revista Brasileira de Zoologia*, 14(1): 99-112.
- Martins U.R. & Galileo M.H.M., 1998. – Gêneros sul-americanos de Desmiphorini (Coleoptera, Cerambycidae) com lados do protórax desarmados. *Revista Brasileira de Entomologia*, 41(2-4): 257-265.
- Martins U.R. & Galileo M.H.M., 2006. – Calliini (Coleoptera, Cerambycidae, Lamiinae): descrições, homonímia, novo registro e chave para as espécies de *Graminea* Thomson, 1864. *Papéis Avulsos de Zoologia*, 46(18): 211-218.
- Martins U.R. & Galileo M.H.M., 2013. – Cerambycidae (Coleoptera) do Museu de Zoologia da Universidade Estadual de Feira de Santana, Bahia. *Iheringia, Série Zoologia*, 103(3): 318-328.
- Martins U.R. & Galileo M.H.M., 2014. – Subfamília Lamiinae, Hemilophini Thomson, 1868, pp. 6–231. Parte I. In: Martins, U.R. (Org.), *Cerambycidae Sul-Americanos (Coleoptera) Taxonomia*. Vol. 13. Sociedade Brasileira de Entomologia, Curitiba, 231 pp.
- Martins U.R., Galileo M.H.M. & Limeira-de-Oliveira F., 2011. – Cerambycidae (Coleoptera) do Estado de Maranhão, Brasil III. *Papéis Avulsos de Zoologia*, 51(18): 275-293.
- MCZbase, 2021. – The Database of the Zoological Collections. Museum of Comparative Zoology – Harvard University. Available from: https://mczbase.mcz.harvard.edu/specimen_images/entomology/large/MCZ-ENT00019723_Asemolea_setosa_hal.jpg (Last accessed 24 February, 2022.)
- Monné M.A., 1980a. – Contribuição ao conhecimento dos Compsomatini (Coleoptera, Cerambycidae, Lamiinae). Parte I. *Revista Brasileira de Entomologia*, 24(1): 29-51.
- Monné M.A., 1980b. – Contribuição ao conhecimento dos Compsomatini (Coleoptera, Cerambycidae, Lamiinae). Parte III. *Revista Brasileira de Entomologia*, 24(3-4): 227-265.
- Monné M.A., 1994. – *Catalogue of the Cerambycidae (Coleoptera) of the Western Hemisphere. Part XVI. Subfamily Lamiinae: Tribes Pogonocherini, Compsomatini, Phacellini, Megabasini and Desmiphorini*. Sociedade Brasileira de Entomologia, São Paulo, 98 pp.
- Monné M.A., 1995. – *Catalogue of the Cerambycidae (Coleoptera) of the Western Hemisphere. Part XX. Subfamily Lamiinae: Tribes Hemilophini, Aerenicini, Pretiliini, Falsamblesthiini and Calliini*. Sociedade Brasileira de Entomologia, São Paulo, 120 pp.
- Monné M.A., 2002. – Catalogue of the Neotropical Cerambycidae (Coleoptera) with known host plant - Part IV: Subfamily Lamiinae, Tribes Batocerini to Xenofreini. *Publicações Avulsas do Museu Nacional*, 94: 1-92.
- Monné M.A., 2005. – Catalogue of the Cerambycidae (Coleoptera) of the Neotropical Region. Part II. Subfamily Lamiinae. *Zootaxa*, 1023: 1-759.
- Monné M.A., 2012. – Catalogue of the type-species of the genera of the Cerambycidae, Disteniidae, Oxypeltidae and Vesperidae (Coleoptera) of the Neotropical Region. *Zootaxa*, 3213: 1-183.
- Monné M.A., 2022. – Catalogue of the Cerambycidae (Coleoptera) of the Neotropical region. Part II. Subfamily Lamiinae. Available from: <https://cerambycids.com/catalog/> (Last accessed 24 February 2022.)
- Monné M.A. & Giesbert E.F., 1994. – Checklist of the Cerambycidae and Disteniidae (Coleoptera) of the Western Hemisphere. Wolfsgarden Books, Burbank, 409 pp.
- Monné M.A. & Hovore F.T., 2006. – A Checklist of the Cerambycidae, or longhorned wood-boring beetles, of the Western Hemisphere. Rancho Dominguez, Bio Quip Publications, 393 pp.
- Monné M.A., Almeida L.M., Oliveira M.L., Viana J.H. & Monné M.L., 2017. – Checklist of Cerambycidae (Coleoptera) primary types of the Coleção Entomológica Pe. Jesus Santiago Moure, Departamento de Zoologia, Universidade Federal do Paraná, Curitiba, Paraná, Brazil, Instituto Nacional de Pesquisas da Amazônia, Amazonas, Manaus, Brazil, and of the Museu Paraense Emílio Goeldi, Pará, Belém, Brazil. *Zootaxa*, 4221(3): 341-365.
- Morvan O. & Morati J., 2006. – Contribution à la connaissance des Cerambycidae de la montagne de Kaw, Guyane Française (Coleoptera). *Lambillionea*, 106(3 – suppl. 2): 3-63.
- Morvan O. & Roguet J-P., 2013. – Inventaire des Cerambycidae de Guyane (Coleoptera). *Supplément au Bulletin de liaison d'ACOREP France "Le Coleopteriste"*, 7: 3-44.
- Moura L.A. & Von Groll E., 2017. – Catalogue of Coleoptera type specimens housed in the collection of the Museu de Ciências Naturais, Fundação Zoobotânica do Rio Grande do Sul, Brazil. *Zootaxa*, 4318(3): 439-473.
- Nascimento F.E.L., Ferreira A.S. & Bravo F., 2017. – Cerambycidae (Coleoptera) do Semiárido: Ampliando o conhecimento. In: Bravo, F. (Org.), *Artrópodes do Semiárido II: Biodiversidade e conservação*. Métais Produção Editorial, São Paulo, 139 pp.
- Noguera F.A. & Chemsak J.A., 1996. – Cerambycidae (Coleoptera), pp. 381–409. In: Llorente Bousquets, J.E. (Ed.), *Biodiversidad taxonomía, y biogeografía de artrópodos de México: Hacia una síntesis de su conocimiento*. Volumen I. Universidad Nacional Autónoma de México, Mexico City, 660 pp.
- Ramírez Hernandez J.J., Santos-Silva A. & Nascimento F.E.L., 2019. – New species, new genera, and new records in Cerambycidae from Peru (Coleoptera). *Annales de la Société entomologique de France* (N.S.), doi.org/10.1080/00379271.2018.1556118: 1-13.
- Roguet J-P., 2021. – Lamiinae. Available from: <https://lamiinae.org/> (Last accessed 18 December 2021.)
- Santos-Silva A., Nascimento F.E.L., Drumont A. & Kozlov A.O., 2019. – Descriptions, notes and new records in South American Cerambycidae (Coleoptera). *Papéis Avulsos de Zoologia*, v.59: e20195915. <http://doi.org/10.11606/1807-0205/2019.59.15>
- Santos-Silva A., Wappes J.E. & Galileo M.H.M., 2018. – Descriptions and synonymies in American Desmiphorini (Coleoptera, Cerambycidae, Lamiinae). *Zootaxa*, 4375(4): 451-501.
- Selander B.S. & Vaurie P., 1962. – A gazetteer to accompany the "Insecta" volumes of the "Biologia Centrali-Americana." *American Museum Novitates*, 2099: 1-70.
- Tavakilian G.L. & Chevillotte H., 2021. – Titan: base de données internationales sur les Cerambycidae ou Longicornes. Available from: <http://titan.gbif.fr/> (Last accessed 24 February 2022.)
- Tavakilian G.L., Berkov A., Meurer-Grimes B. & Mori S., 1997. – Neotropical tree species and their faunas of xylophagous longicorns (Coleoptera: Cerambycidae) in French Guiana. *The Botanical Review*, 63(4): 303-355.
- Touroult J., Dalens P-H, Brûlé S. & Poirier E., 2010. – Inventaire des longicornes: analyse de l'efficacité des techniques de collecte en Guyane. *Supplément au Bulletin de liaison d'ACOREP-France "Le Coléoptériste"*, 15-33.

Résumé

Bezark L. G., Botero J. P. & Santos-Silva A., 2022. – Nouvelles espèces, transferts et note taxonomique de Lamiinae néotropiques (Coleoptera, Cerambycidae). *Faunitaxys*, 10(19): 1 – 25.

Callia ambigua Bates, 1885 est transférée dans le genre *Asemolea* Bates, 1881 (Calliini) dont les caractéristiques morphologiques sont discutées et développées. Des caractéristiques morphologiques supplémentaires sont fournies pour le genre *Murupi* Martins & Galileo, 1998 ; *M. linearis* Martins & Galileo, 1998 est nouvellement cité de Guyane française et une variation chromatique est signalée. Six nouvelles espèces sont décrites : *Scythropopsis tysoni* sp. nov. et *Asemolea rosea* sp. nov. du Guatemala, *Inermestola boliviensis* sp. nov. de Bolivie, *Estola linearis* sp. nov. d'Équateur, *Estoloides elongata* sp. nov. et *Eranina monnei* sp. nov. du Panama. L'inclusion d'*Inermestola chiapasensis* Galileo & Moysés, 2013 dans ce genre est remise en question, indiquant la nécessité d'une étude de l'espèce pour corriger l'attribution générique. *Estola compacta* Breuning, 1940 est signalée d'Équateur et du Pérou, et *E. m-flava* Breuning, 1940 est signalée de Trinité-et-Tobago, dans les États brésiliens d'Amapá et de Pará, et un signalement oublié de l'État brésilien de Goiás est donné. Le deuxième spécimen officiellement connu d'*Eupogonius fuscovittatus* Breuning, 1974 est illustré, et une nouvelle citation départementale au Guatemala est fournie. *Adesmus stephanus* Aurivillius, 1900 est transférée dans le genre *Eranina* Monné, 2005, et *Laraesima ochreoapicalis* Breuning, 1973 (Composomatini) est transférée dans le genre *Estolomimus* Breuning, 1940 (Desmiphorini). *Laraesima fuliginea* Bates, 1885 est redécrite.

Mots-clés. – Coleoptera, Cerambycidae, longicorne, Lamiinae, taxonomie, nouvelles espèces, région néotropicale.

Derniers articles publiés

Gaudin J. & Coache A., 2022. – *Macrommatias* nom. nov., un nouveau nom de remplacement pour *Chandleria* Comellini, 1998 (Coleoptera, Staphylinidae), homonyme récent de *Chandleria* Yamaguti, 1959 (Cestoda, Anoplocephalidae). *Faunitaxys*, 10(9) : 1 – 2.

Lourenço W. R. & Velten J., 2022. – The remarkable variability of the genus *Chaerilobuthus* Lourenço & Beigel, 2011 (Scorpiones: Chaerilobuthidae) and description of a new species from Early Cretaceous Burmite. *Faunitaxys*, 10(10): 1 – 6.

Degallier N. & Tishechkin A.K., 2022. – Révision du genre *Scapicoelis* Marseul, 1862, avec la description de 28 espèces nouvelles (Insecta, Coleoptera, Histeridae, Haeteriinae). *Faunitaxys*, 10(11) : 1 – 87.

Montreuil O. & Uliana M., 2022. – Un nouvel *Amphimallon* Latreille (Coleoptera, Melolonthidae, Rhizotrogini) de l'île de Sifnos (Grèce). *Faunitaxys*, 10(12) : 1 – 5.

Háva J., 2022. – *Anthrenus* (*Anthrenus*) *coacheorum* sp. nov. from Senegal (Coleoptera: Dermestidae: Megatominae). *Faunitaxys*, 10(13) : 1 – 3.

Keith D., 2022. – Description d'une nouvelle espèce du genre *Brachyllus* Brenske, 1896 (Coleoptera, Scarabaeidae, Melolonthinae) de Chine méridionale. *Faunitaxys*, 10(14) : 1 – 3.

Háva J., 2022. – A new Dermestidae species (Coleoptera: Bostrichoidea) from central Iran. *Faunitaxys*, 10(15) : 1 – 3

Coache A. & Borovec R., 2022. – On the genus *Dicasticus* Pascoe, 1886 in archipel of São Tomé and Príncipe (Curculionidae, Entiminae, Peritelini). *Faunitaxys*, 10(16): 1 – 16.

Ballerio A. & Coache A., 2022. – A new species of *Chaetophilharmostes* from São Tomé (Guinea Gulf) with remarks on the generic status of the genus *Chaetophilharmostes* (Coleoptera, Scarabaeoidea, Hybosoridae, Ceratocanthinae). *Faunitaxys*, 10(17): 1 – 8.

Botero J. P. & Santos-Silva A., 2022. – A new species of *Elytrimitatrix* (*Grossifemora*) Santos-Silva & Hovore (Coleoptera, Disteniidae, Disteniinae). *Faunitaxys*, 10(18): 1 – 4.

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- Bulletin of Insectology (Italie)
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- Klapalekiana (République Tchèque)
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- Memorie del Museo Civico di Storia Naturale di Verona (Italie)
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- Proceedings of the Entomological Society of Washington (USA)
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- Stuttgarter Beiträge zur Naturkunde A, Biologie (Allemagne)
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Illustration de la couverture : The first author in a pack train heading into the Darien, southern Panama.

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