

Faunitaxys

*Revue de Faunistique, Taxonomie et Systématique
morphologique et moléculaire*



Volume 7
Numéro 17

Novembre 2019

ISSN: 2269 - 6016
Dépôt légal: Novembre 2019

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morphologique et moléculaire*

ZooBank : <http://zoobank.org/79A36B2E-F645-4F9A-AE2B-ED32CE6771CC>

Directeur de la publication, rédacteur, conception graphique et PAO:

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Elle est disponible librement au téléchargement à partir du site:

<http://faunitaxys.fr/>

La parution de *Faunitaxys* est apériodique

Impression

SPEED COPIE, 6, rue Tréfilerie, F- 42100 Saint-Etienne

Imprimé le 21 novembre 2019

New species and notes on Hexoplonini and Neoibidionini (Coleoptera, Cerambycidae, Cerambycinae)

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Keywords:

Coleoptera ;
Cerambycidae ;
Hexoplonini ;
Neoibidionini ;
Neocompsa ;
Gnomibidion ;
longhorned woodboring beetles ;
Taxonomy ;
Costa Rica ;
Mexico ;
Panama ;
Central America.

Abstract. – Three new species are described: *Compsibidion tuberculatum* n. sp., from Costa Rica; *Neocompsa monnei* n. sp., from Mexico, and *Gnomibidion contiguum* n. sp., from Panama. The male of *Neocompsa wappesi* Giesbert, 1998 is redescribed, and a new country record for Mexico is provided. Chromatic variation in *Heterachthes w-notatus* Linsley, 1935 is reported, and a new country record for Guatemala is provided. Hexoplonini Bouchard *et al.*, 2011 is considered an unjustified emendation for Hexoplonini Martins, 2006. *Notosphaeridion umbrinum* Martins, 1971 is transferred to *Hexoplon* Thomson, 1864. Chromatic variation in *Gnomidolon sylvorum* (Bates, 1892) is reported, and a new state record is provided. The depository institution of the holotypes of *Tropidion hispidum* Martins, 1971, and *Notosphaeridion umbrinum* is corrected.

Bezark L. G. & Santos-Silva A., 2019. – New species and notes on Hexoplonini and Neoibidionini (Coleoptera, Cerambycidae, Cerambycinae). *Faunitaxys*, 7(17): 1 – 17.

ZooBank : <http://zoobank.org/ED664A68-4F9B-4ABC-8043-942F566B3AE0>

Introduction

Over the past several years, the first author was sent a number of specimens of Cerambycidae from Mexico and Central America for study. Examination of the Hexoplonini and Ibidionini allowed us to recognize three new species, redescribe the male of one species, as well as to establish some new distributional records, and to provide comments on several known species.

These two large tribes contain many genera (Hexoplonini, 22 genera and slightly over 150 species; and Neoibidionini, 57 genera with over 500 species). In many cases the distinctions among genera and species in these groups are not always clear and consistent. Herein, we attempt to clarify some of these inconsistencies, and place species in appropriate genera.

Materials and Methods

Photographs were taken with a Canon EOS Rebel T3i DSLR camera, Canon MP-E 65mm f/2.8 1-5X macro lens, controlled by Zerene Stacker AutoMontage software. Measurements were taken in “mm” using measuring ocular Hensoldt/Wetzlar - Mess 10 in the Leica MZ6 stereomicroscope, also used in the study of the specimens.

The collection acronyms used in this study are as follows:

- CASC: California Academy of Sciences, San Francisco, California, USA
- LGBC: Larry G. Bezark, collection, Sacramento, California, USA
- MZSP: Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil
- SMFD: Forschungsinstitut und Naturmuseum Senckenberg, Frankfurt-am-Main, Germany
- ZSMC: Zoologische Staatssammlung des Bayerischen Staates, München, Germany

Taxonomy

NEOIBIDIONINI Monné, 2012

NEOIBIDIONINA Monné, 2012

Compsibidion tuberculatum n. sp.

(Figs. 1-7)

ZooBank : <http://zoobank.org/4DCD9B25-871E-40FE-9954-7BDD9F06DF84>

Holotype, ♂: COSTA RICA, *Guanacaste*: 3 km SE R. Naranjo, 25.I-5.II.1992, F. D. Parker col. (CASC, formerly LGBC).

Paratype, ♀: same data as holotype except 7.V.1992 (LGBC).

Description of the male (Figs. 1-6)

Coloration. – Head and prothorax mostly dark brown, almost black on head; mouthparts reddish-brown except yellowish-brown palpomeres; anterior margin of prothorax and central tubercle brown, and posterior area of prosternal process orangish-brown; mandibles reddish-brown except black margins and apex. Antennae reddish-brown, gradually orangish-brown toward apex. Anterior half of ventral surface of mesothorax dark brown and posterior half reddish-brown, lighter toward apex of mesoventral process. Ventral surface of metathorax mostly dark brown, with anterior process dark reddish-brown, and center of posterior area orangish-brown; elytra mostly brown except pale yellowish-brown posterior seventh and narrow area along suture from anterior quarter, reddish-brown humerus, and epipleural margin reddish-brown on anterior quarter, gradually yellowish-brown toward apex, and two large pale yellow maculae, one oblique, L-shaped on anterior third, from epipleural margin to middle of dorsal surface, another on middle, transverse, from epipleural to sutural margin. Pro- and mesocoxae yellowish-brown; metacoxae mostly reddish-brown; femora reddish-brown except dark reddish-brown apex; tibiae dark reddish-brown, gradually, slightly lighter toward apex; tarsi reddish-brown, slightly lighter toward distal segments. Abdominal ventrites I–IV mostly dark brown, with irregular dark reddish-brown areas; abdominal ventrite V reddish-brown, slightly lighter toward apex.

Head. – Frons finely, abundantly rugose-punctate, except nearly smooth center longitudinal area; with yellowish-white pubescence not obscuring integument (more whitish depending on light intensity). Vertex with sculpturing and pubescence as on frons (pubescence slightly denser). Area behind eyes gradually tumid toward ventral area of lower eye lobes; finely rugose-punctate on area close to upper eye lobes, gradually finely, moderately sparsely punctate on tumid area behind lower eye lobes; remaining surface of area behind upper eye lobes with sculpturing as on tumid area; remaining surface behind lower eye lobes finely, longitudinally striate toward dorsal surface, smooth toward ventral surface; with yellowish-white pubescence partially obscuring integument on area close to upper eye lobe, with sparse yellowish-white pubescence on tumid area, glabrous close to prothorax; with long, erect yellowish-brown setae close to eye (setae slightly more abundant behind lower eye lobe). Antennal tubercles with sculpturing and pubescence as on frons, except smooth and glabrous apex. Genae finely, somewhat rugose-punctate; with sparse yellowish-white pubescence, except smooth and glabrous apex. Postclypeus with sculpturing and pubescence as on frons close to it, finely, sparsely punctate with a few short yellowish-white setae on wide central area, smooth and glabrous laterally; with one long, erect yellowish-brown seta on each side of wide central area. Labrum coplanar with anteclypeus posteriorly, nearly vertical anteriorly, strongly concave centrally; glabrous posteriorly, with long, erect, sparse yellowish-brown setae anteriorly, longer laterally. Distance between upper eye lobes 0.41 times length of scape (0.28 times distance between outer margins of eyes); in frontal view, distance between lower eye lobes 0.62 times length of scape (0.42 times distance between outer margins of eyes). Antennae 2.4 times elytral length, reaching elytral apex at base of antennomere VIII. Scape subcylindrical, slightly widened toward apex; with yellowish-white pubescence not obscuring integument, except glabrous apex (this area gradually widened from dorsal surface to outer surface); with a few long, erect yellowish-brown setae. Antennomeres not carinate dorsally; pedicel and antennomeres with yellowish-brown pubescence not obscuring integument; pedicel and antennomeres III–V with long, erect yellowish-brown setae ventrally and apex of sides and dorsal surface; antennomeres VI–X with a few long, erect yellowish setae on apex.

Antennal formula (ratio) based on length of antennomere III:

– Scape = 0.60. – Pedicel = 0.15. – IV = 0.75. – V = 0.85. – VI = 0.95. – VII = 0.90. – VIII = 0.90. – IX = 0.80. – X = 0.75. – XI = 0.92.

Thorax. – Prothorax subcylindrical, distinctly longer than wide; sides slightly widened between anterior and posterior constrictions, with small gibbosity centrally. Pronotum densely micropunctate, with some coarse, smooth punctures interspersed; with strongly elevated central tubercle; with yellowish-white pubescence partially obscuring integument (whiter depending on light intensity), except glabrous wide, somewhat V-shaped central band, and nearly glabrous anterolateral areas; with a few long, erect yellowish-brown setae. Sides of prothorax with pubescence and erect setae as on pronotum, except glabrous anterior area (this area gradually widened toward prosternum). Prosternum glabrous on center of posterior half, with yellowish-white pubescence on sides of posterior half (whiter depending on light intensity); glabrous on anterior half. Central area of prosternal process distinctly narrowed. Ventral surface of meso- and metathorax with yellowish-white pubescence (whiter depending on light intensity), denser laterally, nearly absent on mesoventral process and central area of metaventricle; metaventricle with long, erect, sparse yellowish-brown setae; apex of mesoventral process rounded projected laterally. Scutellum with yellowish-white pubescence (whiter depending on light intensity) partially obscuring integument.

Elytra. – Finely, sparsely punctate; with long, erect yellowish-brown setae emerging from some punctures, more abundant on posterior quarter; apex truncate, widely concave.

Legs. – Femora with yellowish-white pubescence not obscuring integument dorsally and laterally, nearly absent ventrally; with a few long, erect yellowish-brown setae dorsally and laterally. Tibiae with yellowish-white pubescence not obscuring integument, with long, erect, moderately abundant yellowish-brown setae.

Abdomen. – Ventrites with yellowish-white pubescence not obscuring integument, with long, erect yellowish-brown setae interspersed. Apex of ventrite V emarginate centrally.

Description of the female (Fig. 7).

– *Antennae* 1.95 times elytral length (missing antennomere XI in left antenna, and pedicel and all antennomeres in right antenna), reaching elytral apex at middle of antennomere VIII. – *Antennomeres* IV–VI brown on posterior half (IV irregularly brown on part of anterior half). – *Base of elytra* reddish-brown. – *Antennomeres* not carinate dorsally.

Dimensions (mm), holotype male/paratype female.

– Total length: 9.90/11.20
 – Prothorax: length, 1.90/2.10
 – Anterior width: 1.20/1.30
 – Posterior width: 1.30/1.35
 – Widest prothoracic width: 1.40/1.55
 – Humeral width: 1.85/2.00
 – Elytral length: 6.10/6.85

Remarks.

– According to Martins (1969) and Martins & Galileo (2007), the antennomeres in females of *Compsibidion* Thomson, 1864 are carinate dorsally.

– The female of *C. tuberculatum* n. sp. however, has non-carinate antennomeres.

– Additionally, there are other exceptions. Females of:

- *C. maculatum* Martins, Galileo & Limeira-de-Oliveira, 2011
- *C. marqueti* Audureau, 2015
- *C. amboensis* Santos-Silva, Galileo & Wappes, 2017
- *C. achiraensis* Santos-Silva, Galileo & Wappes, 2017

have non-carinate antennomeres.

– *C. tuberculatum* n. sp. differs from *C. maculatum* as follows:

- pronotum with distinctly elevated central tubercle (absent in *C. maculatum*)
- pronotum mostly pubescent (pubescent only on posterior area in *C. maculatum*)
- elytral apex without spine at outer angle (present in *C. maculatum*)

– *C. tuberculatum* n. sp. differs from *C. ilium* (Thomson, 1864) and *C. quadrisignatum* (Thomson, 1865) by:

- non-carinate antennomeres (carinate in *C. ilium* and *C. quadrisignatum*)
- elytral apex not spiniform (spiniform in *C. ilium* and *C. quadrisignatum*)
- posterior pale yellow macula of the elytra transverse (elliptical and oblique in *C. ilium* and *C. quadrisignatum*)

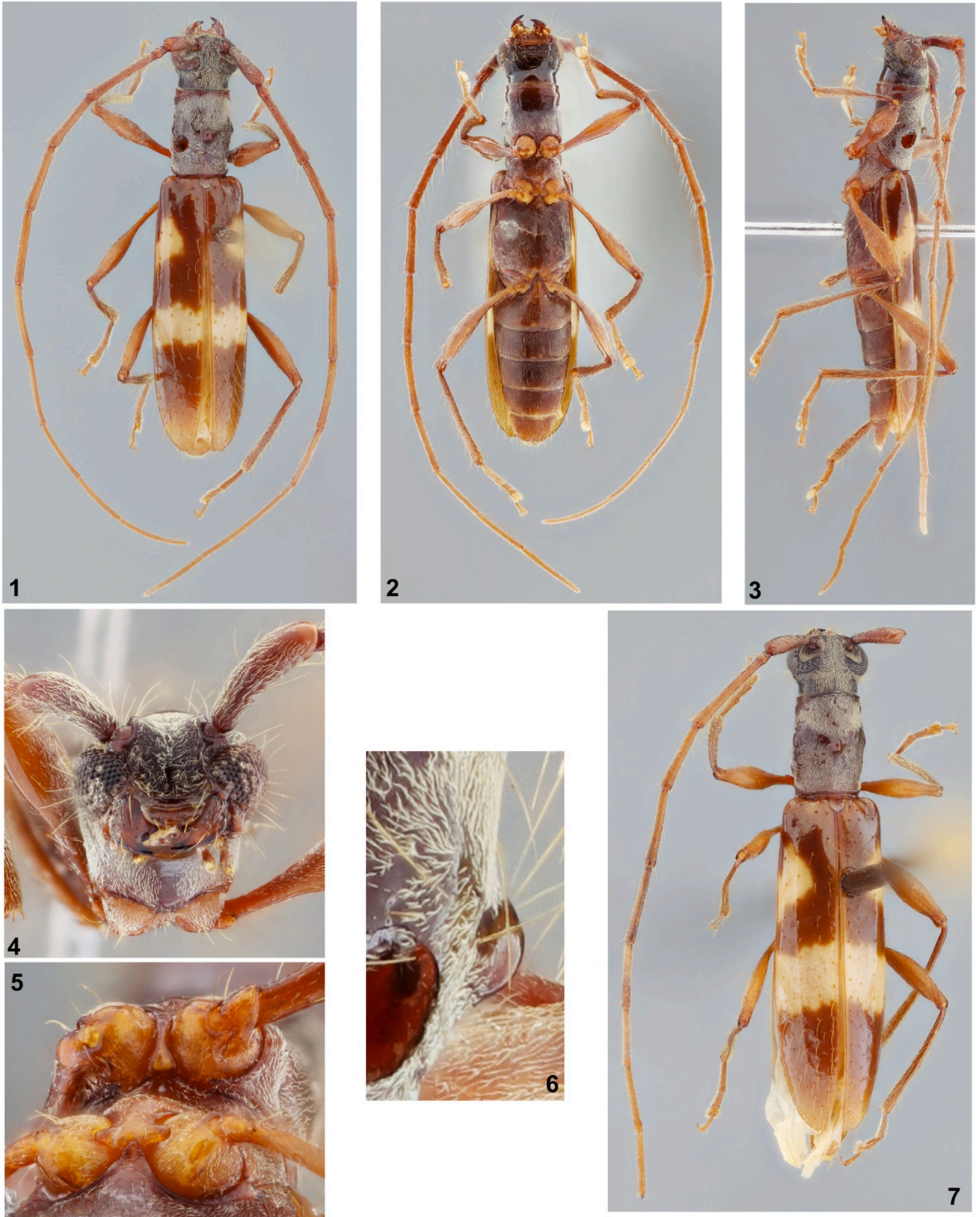
– *C. tuberculatum* n. sp. differs from *C. triviale* Napp & Martins, 1985 by:

- non-carinate antennomeres (carinate in *C. triviale*)
- pronotum with central tubercle distinctly elevated (absent in *C. triviale*)

Etymology. – The specific epithet *tuberculatum* refers to the distinct tubercle on the pronotum which separates this species from several of its congeners.

C. tuberculatum n. sp. can be included in the alternative of couplet “4” from Santos-Silva *et al.* (2017) (modified):

- 4(2). Upper eye lobes with 5 rows of ommatidia; elytra at least partially with moderately abundant, decumbent pubescence.
Bolivia *C. amboensis* Santos-Silva, Galileo & Wappes, 2017
 — Upper eye lobes with 3 or 4 rows of ommatidia; elytra with very sparse, decumbent pubescence or without decumbent pubescence 4'
- 4'(4). Central tubercle of the pronotum slightly elevated; elytral apex rounded; elytra with a single pale yellow macula.
Bolivia *C. achiraensis* Santos-Silva, Galileo & Wappes, 2017
 — Central tubercle of the pronotum strongly elevated; elytral apex truncate; elytra with two pale yellow maculae.
Costa Rica *C. tuberculatum* Bezark & Santos-Silva n. sp.



Figures 1-7. *Compsibidion tuberculatum* n. sp. 1-6) Holotype ♂: 1) Dorsal habitus; 2) Ventral habitus; 3) Lateral habitus; 4) Head, frontal view; 5) Mesothorax, ventral view; 6) Pronotum, lateral view. 7) Paratype ♀, dorsal habitus.

COMPSINA Martins & Galileo, 2007***Heterachthes w-notatus*** Linsley, 1935

(Figs. 8–10)

Heterachthes w-notatus Linsley, 1935: 80;
 Blackwelder, 1946: 570 (checklist);
 Gibson & Carrillo, 1959: 118 (distr.);
 Martins & Chemsak, 1966: 459;
 Martins, 1970: 1305;
 Chemsak *et al.*, 1992: 51 (checklist);
 Monné, 1993: 79 (cat.);
 Monné & Giesbert, 1994: 79 (checklist);
 Noguera & Chemsak, 1996: 399 (checklist);
 Noguera *et al.*, 2002: 624 (distr.);
 Monné, 2005: 375 (cat.);
 Monné & Hovore, 2006: 98 (checklist);
 MacRae *et al.*, 2012: 178 (hosts);
 Noguera *et al.*, 2012: 620 (distr.);
 Bezark, 2013: 45 (distr.);
 Heffern *et al.*, 2018: figs. 9–10;
 Bezark, 2019a: 131 (checklist); 2019b: 408 (distr.);
 Monné, 2019: 538 (cat.).

Remarks

– *Heterachthes w-notatus* Linsley, 1935 was described from Mexico (Mexico). Currently, it is known from the Mexican states of Sonora, Nayarit, Michoacán, Morelos, Puebla, Mexico, Oaxaca, Veracruz and Jalisco (Monné 2019).

– The female (Fig. 8) examined from Guatemala is darker than nearly all females from Mexico. The female from Guatemala (Fig. 8) and one female from Mexico (Oaxaca) (Fig. 9) differ from the typical form of the species (Fig. 10) by the pale yellow band on elytra not W-shaped. However, there are no other morphological differences between all specimens examined. The female from Guatemala (Fig. 8) has the antennae dark brown (mostly yellowish-brown to reddish-brown in the other specimens) and erect setae yellowish (white in the other specimens). Even so we believe that these are only specific variations, which appears to be more evident by the existence of a female with intermediate color (Fig. 9): elytral pale yellow band similar to that in the female from Guatemala (Fig. 8), and the remaining general color as in the typical form (Fig. 10).

Material examined

– MEXICO, *Oaxaca*: vic. El Camarón, 1 km S Hwy 190, Rd to Microondas San Cristobal, 4112', 1 ♀, 26.VI.2014, J. Rifkind col. (LGBC).

– GUATEMALA (**New country record**), *Zacapa*: Estación Biológica Heloderma near El Arenal, 546 m, 14.86321°N / 89.78526°W, 1 female, 2.VI.2016, R.S. Zack col. (LGBC).

Neocompsa wappesi Giesbert, 1998

(Figs. 19–22)

Neocompsa wappesi Giesbert, 1998: 47;
 Turnbow *et al.*, 2003: 12 (distr.);
 Monné, 2005: 385 (cat.);
 Monné & Hovore, 2006: 99 (checklist);
 Hovore, 2006: 373 (distr.);
 Monné, 2019: 554 (cat.);
 Bezark, 2019a: 132 (checklist).

Redescription of the male

Coloration. – Integument mostly dark brown; mouthparts reddish-brown, more yellowish on apex of palpomeres; antennal tubercles and clypeus mostly dark reddish-brown; labrum and genae dark reddish-brown posteriorly, yellowish-brown anteriorly; basal 2/3 of mandibles dark reddish-brown, except black margins, and distal third black; scape dark

brown basally, gradually dark reddish-brown toward apex; pedicel dark brown on base, orangish-brown on remaining surface; antennomeres III–VI yellowish-brown, more brownish on apex, and dark brown on dorsal carina of basal antennomeres; remaining antennomeres orangish-brown, with apex of VII–X slightly darkened; prosternal process and mesoventral process reddish-brown toward apex. Elytra gradually from reddish-brown to orangish-brown along suture; humeri dark reddish-brown; each elytron with two pale yellow maculae: one longitudinal, subelliptical on anterior third of dorsal surface; another oblique on posterior third, from epipleural to sutural margin. Coxae partially reddish-brown; peduncle of femora light yellowish-brown; tibiae light yellowish-brown on anterior 2/3, brown on posterior third; tarsi mostly light brown.

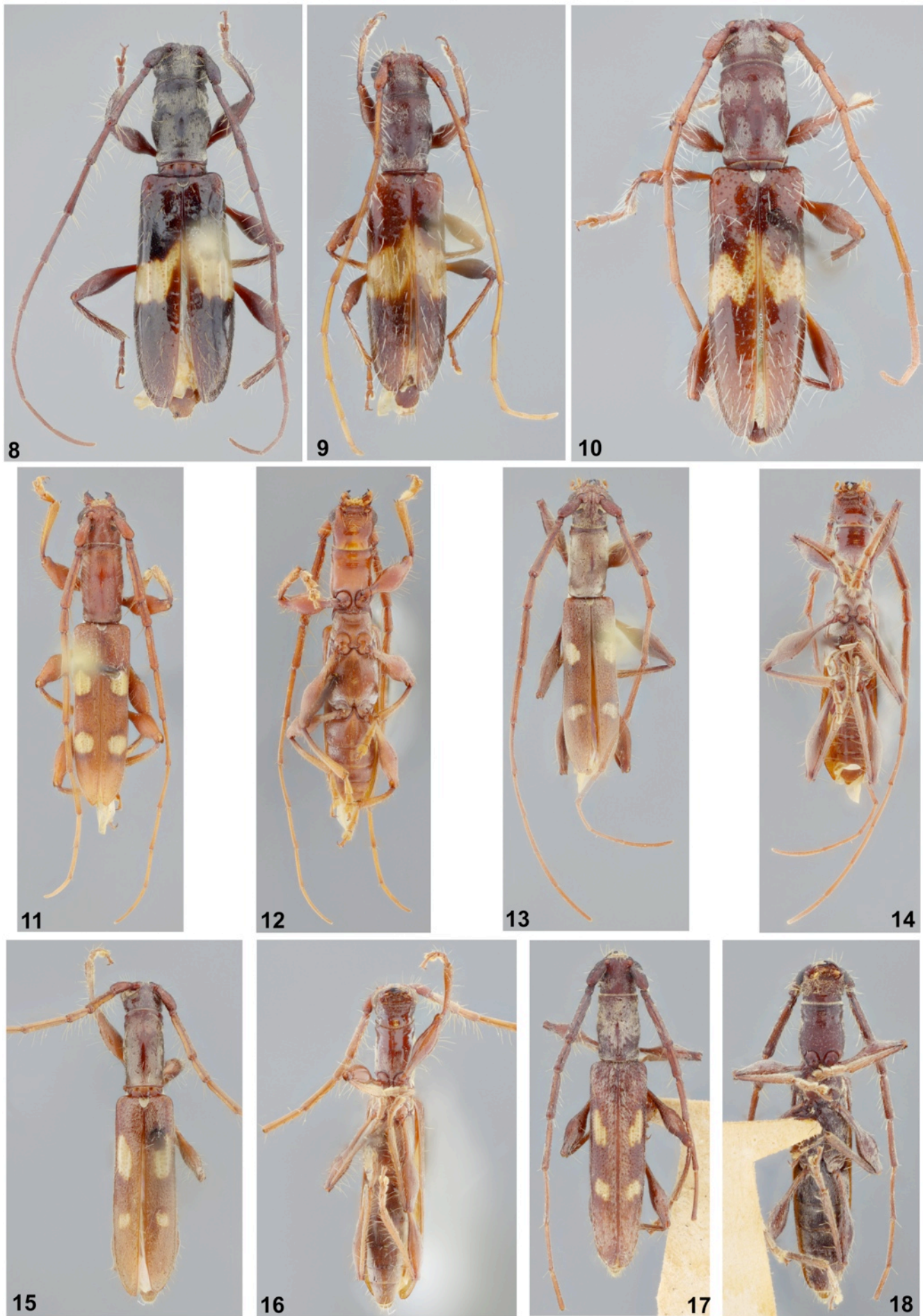
Head. – Frontal plate finely striate laterally, smooth centrally; median groove deep on frons; remaining surface of frons finely, abundantly punctate; with short yellowish-white pubescence not obscuring integument (whiter depending on light intensity), absent on smooth area of frontal plate. Vertex finely, abundantly punctate from area between antennal tubercles to middle of area between eyes and prothorax, finely, transversely rugose on remaining surface; with yellowish-white pubescence partially obscuring integument on finely punctate area, glabrous on rugose area; with a few long, erect yellowish setae. Area behind eyes with sculpturing, pubescence and erect setae as on vertex on area close to eye, smooth, glabrous on area close to prothorax. Antennal tubercles with sculpturing and pubescence as on frons on wide basal region, smooth, glabrous on apex. Area between eyes and antennal sockets with dense yellowish-brown pubescence. Genae finely, sparsely punctate, with sparse yellowish-brown pubescence close to eye, smooth, glabrous on apex. Postclypeus with yellowish-brown pubescence close to frons, nearly obscuring integument, sparse, yellowish-white anteriorly, except glabrous sides; with one long, erect yellowish-brown seta on each side of wide central area. Labrum with long, erect, moderately abundant yellowish-brown setae on wide anteroventral area (setae longer laterally). Distance between upper eye lobes 0.6 times length of scape (0.37 times distance between outer margins of eyes); in frontal view, distance between lower eye lobes 0.8 times length of scape (0.5 times distance between outer margins of eyes). Antennae 2.5 times elytral length, reaching elytral apex at apex of antennomere VII. Scape with yellowish-white pubescence, sparser ventrally, absent on outer apex of dorsal surface, and apex of outer surface; with a few long, erect yellowish setae ventrally and laterally. Pedicel with sparse yellowish-white pubescence except glabrous apex; with a few long, erect yellowish setae. Dorsal carina of antennomeres III–V well marked; ventral surface of antennomeres III–V with long, erect yellowish setae; apex of antennomeres III–X with moderately long, erect yellowish setae.

Antennal formula (ratio) based on length of antennomere III:

– Scape = 0.49. – Pedicel = 0.13. – IV = 0.60. – V = 0.91. – VI = 0.97. – VII = 0.89. – VIII = 0.87. – IX = 0.78. – X = 0.78. – XI = 0.87.

Thorax. – Prothorax cylindrical, distinctly longer than wide; parallel-sided, anterior and posterior constrictions slightly distinct dorsally. Pronotum slightly longitudinally carinate centrally from base to apex; with four small tubercles, one on each side of apex of anterior third, another on each side of posterior quarter; coarsely, sparsely punctate; with yellowish-white pubescence partially obscuring integument, absent on longitudinal central area, and most of area close to anterior margin (this latter with narrow transverse pubescent band centrally); with long, erect, sparse yellowish setae. Sides of prothorax coarsely sparsely punctate, except smooth anterior area (this latter gradually widened toward prosternum); with yellowish-white pubescence partially obscuring integument, except nearly glabrous smooth area; with a few long, erect yellowish setae. Prosternum with yellowish-white pubescence partially obscuring integument on posterior half, nearly glabrous on anterior half; narrowest area of prosternal process 0.2 times width of procoxal cavity. Ventral surface of meso- and metathorax with yellowish-white pubescence, denser on mesothorax and sides of metathorax; with a few long, erect yellowish setae on metathorax. Scutellum with dense yellowish-white pubescence.

Elytra. – Coarsely, moderately abundant punctate on anterior 2/4, finer sparser on posterior quarter; with short yellowish-white



Figures 8-18. 8-10) *Heterachthes w-notatus*, ♀, dorsal habitus: 8) Specimen from Guatemala; 9) Specimen from Mexico, Oaxaca; 10) Specimen from Mexico, Michoacán. 11-12) *Neocompsa macrotricha*, ♂ from Mexico (Oaxaca): 11) Dorsal habitus; 12) Ventral habitus. 13-14) *Neocompsa macroscina*, ♂ from Mexico (Jalisco): 13) Dorsal habitus; 14) Ventral habitus. 15-16) *Neocompsa agnosta*, paratype ♂ from Mexico (Nayarit): 15) Dorsal habitus; 16) Ventral habitus. 17-18) *Neocompsa quadriplagiata*, ♂ from Mexico: 17) Dorsal habitus; 18) Ventral habitus.

pubescence not obscuring integument throughout (including on pale yellow maculae); apex slightly obliquely truncate.

Legs. – Femora and tibiae with yellowish-white pubescence not obscuring integument, with long, erect yellowish setae interspersed; apex of femoral club distinctly narrowed, especially metafemora; pro- and mesofemora with carina on lateral apex, absent on metafemora.

Abdomen. – Ventrites with yellowish-white pubescence partially obscuring integument, with a few long, erect yellowish setae interspersed (slightly denser toward distal segments). Apex of ventrite V slightly concave.

Dimensions (mm), male

- Total length: 10.00
- Prothorax, length: 2.30
- Prothorax, anterior width: 1.15
- Prothorax, posterior width: 1.20
- Widest prothoracic width: 1.30
- Humeral width, 1.80
- Elytral length, 6.45.

Material examined

– MEXICO (**New country record**), *Chiapas*: Hwy 190, 12.3–16.3 km N Arriaga, “La Sepultura”, 20.VI.2016, T.D.F., J. Rifkind & E. Martinez col. (LGBC).

Remarks

Neocompsa wappesi was described and remains known from Guatemala and Honduras (Monné 2019). Males of *Neocompsa wappesi* differ from those of *N. macrotricha* Martins, 1970 (Figs. 11–12) as follows: antennae longer, reaching elytral apex at apex of antennomere VII; antennomere IV as long as width of prothorax; pubescence of the prosternum covers the entire posterior half; apex of femora narrower; femora bicolorous; peduncle of femora longer and more slender. In *N. macrotricha*, antennae reaching elytral apex at middle of antennomere VIII, antennomere IV distinctly shorter than width of prothorax, pubescence of the prosternum restricted to area near procoxal cavities, apex of femora wider, femora unicolorous, and peduncle of femora shorter and wider. Males of *Neocompsa wappesi* differ from males of *N. macroscina* Martins, 1970 (Figs. 13–14) by the femora more slender and bicolorous (stouter and unicolorous in *N. macroscina*), and antennomeres distinctly lighter (darker in *N. macroscina*). It differs from males of *N. quadriplagiata* (LeConte, 1873) (Figs. 17–18) by the posterior elytral pale yellow macula placed more near apex (posterior quarter), and ascendant from epipleural to sutural margin (about base of posterior third, transverse or descendent toward apex, not reaching epipleura and suture in *N. quadriplagiata*). Male *Neocompsa wappesi* differs from males of *N. agnosta* Martins, 1970 (Figs. 15–16) by the anterior pale yellow macula on elytra pubescent (glabrous in *N. agnosta*), and antennomere IV as long as prothoracic width (shorter than prothoracic width in *N. agnosta*). It differs from *N. textilis* (Thomson, 1865) (Figs. 23–24) by the elytra truncate at apex (spiniform in *N. textilis*), and antennal tubercles more distant from each other (closer in *N. textilis*). *Neocompsa wappesi* differs from *N. giesberti* Martins & Napp, 1986 (see photograph of the holotype at Bezark 2019c) by the presence of tubercles on pronotum (absent in *N. giesberti*); from *N. intricata* Martins, 1970 (see photograph of the holotype at Bezark 2019c) by the elytral apex lacking projection on outer angle (outer angle with short spine in *N. intricata*); from *N. alacris* (Bates, 1885) (see photograph of the holotype at Bezark 2019c) by the elytra entirely pubescent (lacking pubescence on basal region of the elytra in *N. alacris*), and by the antennomere IV as long as prothoracic width (shorter in *N. alacris*); from *N. eburioides* (Thomson, 1867) (see photograph of the holotype at

Bezark 2019c) by the anterior pale yellow macula of the elytra pubescent (not so in *N. eburioides*), and elytral apex without spine (not pubescent and with spine at outer angle in *N. eburioides*); from *N. glaphyra* Martins, 1970 (see photograph of the holotype at Bezark 2019c) by the elytra pubescent (not so in *N. glaphyra*); from *N. habra* Martins, 1970 (see photograph of the holotype at Bezark 2019) by the anterior pale yellow macula of the elytra pubescent (not so in *N. habra*); from *N. leechi* Martins, 1970 (see photograph of the holotype at Bezark 2019c) by the elytral punctures not contrasting (contrasting in *N. leechi*), and by the anterior half of the prosternum not pubescent (pubescent in *N. leechi*); from *N. lenticula* Martins (1970) (see photograph of the holotype at Bezark 2019c) by the elytra pubescent (not so in *N. lenticula*); from *N. obscura* Martins, 2009 (see photograph of the holotype at Bezark 2019c) by the posterior pale yellow macula of the elytra oblique, from epipleural to sutural margin (dorsal and elliptical in *N. obscura*); from *N. pallida* Martins & Galileo, 2010 (see photograph of the holotype at Bezark 2019c) by the elytral outer apex not projected (spiniform in *N. pallida*); from *N. spinosa* Martins, 1970 (see photograph of the holotype at Bezark 2019c) by the elytral outer apex not projected (spiniform in *N. spinosa*); from *N. squalida* (Thomson, 1867) (see photograph at Bezark 2019c) by the elytral pale yellow maculae not rounded (rounded in *N. squalida*), and outer angle of the elytra not spiniform (spiniform in *N. squalida*); from *N. tuberosa* Martins, 1970 (see photograph of the holotype at Bezark 2019c) by the elytra entirely pubescent (not pubescent anteriorly in *N. tuberosa*); and from *N. ventricosa* (Bates, 1885) (see photograph of the lectotype at Bezark 2019c) by the outer angle of the elytra not spiniform (spiniform in *N. ventricosa*).

***Neocompsa monnei* n. sp.**

(Figs. 25–33)

ZooBank : <http://zoobank.org/B666FBEE-0B5F-4E3C-BE57-1A22AC37BC01>

Holotype, ♂: MEXICO, *Oaxaca*: Hwy 190, 5 km W junction road to San Dionisio, Ocoatepec, 2752', dry Oak forest, 24.VI.2014, J. Rifkind col. (CASC, formerly LGBC).

Paratypes

- 2 ♂, 4 ♀, same data as holotype (1 ♂, 3 ♀ LGBC; 1 ♂, 1 ♀, MZSP),
- 1 ♂, 1 ♀, MEXICO, *Oaxaca*: Hwy 135 & road to San Juan Tonaltepec, 16 July, 2018, J. Rifkind & E. Martinez col. (LGBC),
- 1 ♂, MEXICO, *Oaxaca*: 4.6–9 km N Diaz Ordez, 6400–7570', 8 July, 2018 J. Rifkind & E. Martinez col. (LGBC).

Description of the male (Figs. 25–28, 32–33)

Coloration. – Vertex and area behind upper eye lobes black; remaining surface of head reddish-brown; mouthparts reddish-brown, except two last palpomeres dark brown on basal 2/3, yellowish brown on distal third; mandibles reddish-brown on basal 2/3 (margins darkened), black on posterior third; scape, pedicel and antennomeres III–IV dark brown, and remaining antennomeres gradually reddish-brown toward XI (usually, distal area of each antennomere slightly darker); prothorax black on pronotum, and remaining surface gradually dark reddish-brown toward ventral surface; meso- and metathorax reddish-brown; elytra black, except basal area reddish-brown (this area distinctly larger, covering about basal quarter laterally); elytra with narrow, oblique pale yellow band on anterior third; femoral peduncle reddish-brown; femoral club dark brown; tibiae and tarsi dark brown; abdominal ventrites dark brown, with distal area of some ventrites dark reddish-brown. Yellowish-white pubescence appearing to be white or grayish-white depending on light intensity.

Head. – Frontal plate not well-delimited centrally; frontal plate and remaining surface of frons minutely striated, finely, sparsely punctate centrally, partially confluent punctate laterally, smooth on center of



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Figures 19-24. 19-22) *Neocompsa wappesi*, ♂: 19) Dorsal habitus; 20) Ventral habitus; 21) Lateral habitus; 22) Head, frontal view. 23-24) *Neocompsa textilis*, ♂ from Mexico (Yucatán): 23) Dorsal habitus; 24) Ventral habitus.

frontal plate; with short, very sparse yellowish-white setae. Area between antennal tubercles finely, confluent punctate; area from between upper eye lobes to near prothorax, finely, densely, confluent punctate (punctures finer than between antennal tubercles); area of vertex adjacent to prothoracic margin finely, transversely rugose-punctate; area between antennal tubercles and upper eye lobes with short, very sparse yellowish-white setae; remaining confluent punctate region with abundant yellowish-white pubescence; area adjacent to prothorax glabrous; with a few long, erect yellowish setae between upper eye lobes and on confluent punctate area of vertex. Area behind upper eye lobes with sculpturing and pubescence as on vertex between posterior margin of upper eye lobes and prothoracic margin; area behind lower eye lobes moderately coarsely, sparsely punctate on wide area close to eye, finely, sparsely punctate close to prothorax; wide area close to eye with both, short and long, sparse yellowish setae, and remaining surface glabrous. Antennal tubercles minutely striate, with fine punctures interspersed, except smooth central area, and nearly smooth apex; with short, sparse yellowish-white setae, except glabrous smooth area. Genae finely, sparsely punctate along central area, smooth close to eye and on apex; with short, sparse yellowish-white setae on punctate area, glabrous on smooth areas. Postclypeus very finely punctate on wide central area close to frontal plate, smooth on remaining surface; with short, very sparse yellowish setae on wide central area, glabrous on remaining surface, and with one long, erect yellowish setae on each side of wide central area. Labrum smooth and glabrous close to anteclypeus, finely punctate, with long, erect yellow setae anteriorly (longer laterally). Gulamentum glabrous, nearly smooth on posterior third, transversely striate-punctate, with sparse, both short and long yellowish-white setae. Distance between upper eye lobes 0.55 times length of scape (0.35 times distance between outer margins of eyes); in frontal view, distance between lower eye lobes 0.83 times length of scape (0.53 times distance between outer margins of eyes). Antennae 2.7 times elytral length, reaching elytral apex at posterior third of antennomere VII. Scape finely, abundantly punctate except smooth outer side of dorsal surface and posterior area of outer surface; with short, sparse yellowish-white setae, absent on smooth area; with a few short, erect yellowish setae. Pedicel and antennomeres with yellowish-white pubescence not obscuring integument, except glabrous inner ventral surface of basal antennomeres; antennomeres III–V with long, erect yellowish setae (sparser toward V); apex of III–X with long, erect, sparse yellowish setae throughout; basal antennomeres distinctly carinate dorsally.

Antennal formula (ratio) based on length of antennomere III:

– Scape = 0.45. – Pedicel = 0.13. – IV = 0.63. – V = 0.89. – VI = 0.89. – VII = 0.88. – VIII = 0.76. – IX = 0.74. – X = 0.63. – XI = 0.85.

Thorax. – Prothorax cylindrical, distinctly longer than wide; parallel-sided, anterior and posterior constrictions moderately well-marked. Pronotum with slightly elevated, longitudinal gibbosity centrally from just before middle to posterior quarter; densely micropunctate with coarse punctures interspersed, except smooth central gibbosity, narrow central area from anterior margin to central gibbosity, narrow area close to posterior margin, and nearly smooth anterior area; with grayish-white pubescence partially obscuring integument, with long, erect yellowish setae interspersed, except glabrous smooth areas. Sides of prothorax with pubescence, sculpturing and erect setae as on sides of prothorax, except smooth and glabrous anterior and posterior areas, both widened toward prosternum, especially anterior one. Prosternum with sparse grayish-white pubescence close to procoxal cavities, and somewhat U-shaped band on center of posterior half; remaining surface with a few short grayish-white setae; with a few long, erect yellowish setae on posterior third; narrowest area of prosternal process 0.15 times width of procoxal cavity. Ventral surface of mesothorax and sides of metathorax with grayish-white pubescence partially obscuring integument; remaining surface of metathorax with sparse grayish-white pubescence, nearly absent centrally, with long, erect yellowish setae interspersed. Scutellum with dense grayish-white pubescence, denser toward apex.

Elytra. – Coarsely, moderately abundantly punctate, punctures gradually finer and sparser toward apex; apex nearly rounded; with

short, suberect grayish-white setae, sparser basally, gradually more abundant toward posterior fifth, distinctly more abundant on posterior fifth; with long, erect yellowish setae throughout, more abundant from middle.

Legs. – Femora and tibiae with grayish-white pubescence, except ventral surface of club, with long, erect yellowish setae interspersed; profemora with carina on lateral apex, absent on meso- and metafemora.

Abdomen. – Ventrites with grayish-white pubescence, sparser centrally on I–IV, absent on posterocentral area of I–II, and apex of I–IV; with long, erect, sparse yellowish setae interspersed; apex of ventrite V nearly truncate.

Description of the female (Fig. 29–31). – Differs from male by the antennae shorter, 1.9 times elytral length, reaching elytral apex at slightly after middle of antennomere IX, and apex of abdominal ventrite V narrower and distinctly rounded.

Variation. – Only last palpomeres dark brown on basal 2/3 and yellowish brown on distal third; antennae entirely dark brown or with distal segments yellowish-brown with their apex darkened; elytra without pale yellow maculae (Fig. 29); elytra without pale yellow macula anteriorly, with slightly oblique pale yellow macula dorsally after middle (Fig. 30); elytra with pale yellow anterior macula short, and with lightly oblique pale yellow macula dorsally beyond middle (Fig. 31); elytra with pale yellow anterior macula short, without pale yellow macula beyond middle (Fig. 32); vertex with very sparse yellowish-white pubescence throughout (Fig. 30).

Dimensions (mm), holotype male/paratypes male/paratypes female.

– Total length: 9.40/7.45–7.90/8.10–9.40

– Prothorax length: 2.30/1.70–1.80/1.70–2.00

– Prothorax anterior width: 1.15/0.90–0.95/0.95–1.10

– Prothorax posterior width: 1.15/0.85–0.90/0.95–1.10

– Widest prothoracic width: 1.25/0.95–1.00/1.00–1.15

– Humeral width: 1.65/1.30–1.35/1.40–1.65

– Elytral length: 5.80/4.60–4.80/4.95–6.00

Remarks

Neocompsa monnei n. sp. is similar to *N. ptoma* Martins, 1970 (see photograph of the holotype at Bezark 2019c), but differs by the pronotum almost entirely pubescent (only close to anterior and posterior margins in *N. ptoma*), femora light on peduncle, dark on club (femora entirely light or peduncle dark and club light in *N. ptoma*). It differs from *N. sinaloana* (Linsley, 1935) (see photograph of the holotype at Bezark 2019c) by the elytra without abundant pubescence (present in *N. sinaloana*); from *N. tenuissima* (Bates, 1885) (see photograph of the lectotype at Bezark 2019c) by the elytra almost entirely unicolorous (distinctly bicolorous in *N. tenuissima*), and pronotum unicolorous (bicolorous in *N. tenuissima*); from *N. clerochroa* (Thomson, 1867) by the elytra almost entirely unicolorous (distinctly bicolorous in *N. clerochroa*), and pronotum mostly pubescent (not or almost not so in *N. clerochroa*); and from *N. turnbowi* Giesbert, 1998 (see photograph of the holotype at Bezark 2019c) by the elytral apex not distinctly narrowed posteriorly (distinctly narrowed in *N. turnbowi*), and anterior light macula of the elytra, when present, distinctly shorter (longer in *N. turnbowi*).

Etymology. – The species name *monnei* is named for both Miguel & Marcela Monne, who were so gracious to the first author when he visited them and the museum in Rio de Janeiro in 2012.



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Figures 25-33. *Neocompsa monnei* n. sp. 25-28) Holotype ♂: 25) Dorsal habitus; 26) Ventral habitus; 27) Lateral habitus; 28) Head, frontal view. 29-33) Paratypes, dorsal view: 29) Female 1; 30) Female 2; 31) Female 3; 32) Male 1; 33) Male 2.

TROPIDINA Galileo & Martins, 2007

***Gnomibidion contiguum* n. sp.**

(Figs. 34–37)

ZooBank : <http://zoobank.org/B4AAA7A5-C12E-4D4D-BE68-0E18EA165C3E>

Holotype, ♂: PANAMA, *Panama*: 31 km E Canita, 21.V.1991, F.T. Hovore col. (CASC, formerly LGBC).

Paratype, ♂: same data (LGBC).

Description of the male

Coloration. – Integument mostly reddish-brown; mandibles almost black on posterior third and margins of anterior 2/3; scape dark brown basally; pedicel dark brown on basal third; antennomere III dark brown; antennomere IV dark reddish-brown, with irregular brown areas; remaining antennomeres orangish-brown. Elytra reddish-brown basally, with large, elliptical yellow macula from basal fifth to slightly beyond middle, margined by wide dark brown band anteriorly and posteriorly; remaining surface orangish-brown. Ventral surface of metathorax slightly, gradually lighter toward apex. Femora dark brown on peduncle and apex of club, reddish-brown on remaining surface of club (more distinctly so on profemora, and dark area covering large area on base of club); tibiae dark brown, slightly lighter on posterior area. Abdominal ventrites I–III brown on apex; ventrite IV reddish-brown on basal half, gradually orangish-brown on posterior half; ventrite V mostly orangish-brown.

Head. – Frontal plate large, subtriangular, finely, densely striated; area of frons between frontal plate and antennal tubercles slightly depressed, finely striated (striae perpendicular to the striae on frontal plate); with a few minute white setae close to antennal tubercles, glabrous on remaining surface. Area between antennal tubercles and upper eye lobes slightly rugose-punctate; remaining surface of vertex finely, sparsely punctate, except area close to prothoracic margin with denser punctures; with minute, very sparse yellowish-white setae. Area behind upper eye lobes smooth close to eye, finely, sparsely punctate on remaining surface (punctures slightly more abundant close to prothoracic margin); with a few short yellowish setae, and one long, erect yellowish seta close to eye. Area behind lower eye lobes tumid, finely, sparsely punctate close to eye; area closer to prothorax somewhat rugose superiorly, finely sparsely punctate toward ventral surface; with moderately short, erect yellowish setae on inferior area, glabrous on remaining surface. Antennal tubercles moderately elevated, with apex narrowed and blunt; with a few short, white setae. Genae triangularly projected; finely, moderately abundantly punctate; with a few short yellowish setae. Postclypeus very finely striated, with a few coarse punctures interspersed on wide central area, from each emerges one long, erect yellow seta; with a few minute yellowish setae on wide central area, glabrous laterally. Labrum with long, erect yellow and yellowish-white setae, longer laterally. Outer side of mandibles longitudinally rugose-punctate, with long, erect yellow setae on anterior 2/3, smooth, glabrous on posterior third. Gulamentum smooth, glabrous on posterior 2/3 (except a few long, erect setae close to anterior third); anterior third coarsely rugose-punctate, with sparse, both short and long, erect yellowish setae. Distance between upper eye lobes 0.45 times length of scape (0.29 times distance between outer margins of eyes); in frontal view, distance between lower eye lobes 0.69 times length of scape (0.45 times distance between outer margins

of eyes). Antennae 2.9 times elytral length, reaching elytral apex slightly before middle of antennomere VII (reaching elytral apex before apex of antennomere VI in the largest male examined (Fig. 34), in which distal antennomeres are missing). Scape finely, sparsely punctate; with very sparse, minute yellowish-white setae and a few long, yellow setae. Pedicel and antennomeres with sparse yellowish-white pubescence, more abundant toward distal segments; pedicel and antennomeres III–VI with long, erect yellow setae ventrally, more abundant on III; antennomeres III–X with long, erect, sparse yellow setae on apex (shorter than ventral ones).

Antennal formula (ratio) based on length of antennomere III (only smallest specimen measured):

– Scape = 0.47. – Pedicel = 0.16. – IV = 0.77. – V = 0.92. – VI = 0.95. – VII = 0.93. – VIII = 0.84. – IX = 0.77. – X = 0.68. – XI = 0.95.

Thorax. – Prothorax not strongly constricted centrally. Pronotum with slightly elevated central gibbosity, and moderately elevated gibbosity on each side close to posterior constriction; minutely, very sparsely punctate; with a few long, erect yellow setae. Sides of prothorax minutely, very sparsely punctate on wide central area, slightly striated anteriorly and posteriorly; with a few, both long and short yellow setae on wide central area. Prosternum smooth except somewhat rugose area close to widened anterior area; with sparse yellowish-white pubescence close to procoxal cavities, nearly glabrous on remaining surface. Prosternal process with moderately abundant yellowish-white pubescence; narrowest area about 0.12 times width of procoxal cavity. Sides of mesoventrite with sparse yellowish-white pubescence laterally (whiter depending on light intensity), glabrous on remaining surface; mesanepisternum glabrous; mesepimeron and metanepisternum with abundant yellowish-white pubescence (whiter depending on light intensity); metaventrite with sparse yellowish-white pubescence laterally, distinctly denser close to metacoxal cavities (whiter depending on light intensity), and remaining surface with both short and long, erect, sparse yellowish setae. Scutellum with yellowish-white pubescence not obscuring integument (slightly distinct due to the color and size).

Elytra. – Coarsely, moderately abundantly punctate on basal half, finer, sparser with minute, moderately abundant punctures interspersed on posterior half; with minute yellowish-white setae emerging from most punctures, and a few long, erect yellow setae emerging from others; apex emarginate, with distinct projections at outer and sutural angles (outer one longer).

Legs. – Femora with minute, very sparse yellowish-white setae, and long, erect, sparse yellow setae. Tibiae with minute, sparse yellowish-white setae dorsally and laterally, yellow, more abundant ventrally, especially on posterior third, with long, erect yellow setae interspersed.

Abdomen. – Ventrites with short, decumbent, sparse yellowish setae, with long, erect yellow setae interspersed. Apex of ventrite V truncate.

Dimensions (mm), holotype male/paratype female.

– Total length: 11.60–7.80
 – Prothorax length: 2.85–1.85
 – Prothorax anterior width: 1.50–0.90
 – Prothorax posterior width: 1.55–0.95
 – Widest prothoracic width: 1.55–0.95
 – Humeral width, 2.15–1.45
 – Elytral length: 7.10–4.70

Gnomibidion contiguum n. sp. can be included in the alternative of couplet “9” from Martins (1968) (modified):

- | | | |
|--------|---|--|
| 9(1). | Elytra with large yellow macula contrasting with remaining color | 10 |
| — | Elytra lacking yellow macula contrasting with anterior and posterior color. <i>Colombia</i> | <i>G. occultum</i> Martins, 1968 |
| 10(9). | Pronotum with dark macula; outer angle of elytra with long spine. <i>French Guiana, Brazil (Mato Grosso do Sul)</i> | |
| — | | <i>G. translucidum</i> (Martins, 1960) |
| — | Pronotum lacking dark macula; outer angle of elytra with moderately short projection. <i>Panama</i> | |
| — | | <i>G. contiguum</i> Bezark & Santos-Silva n. sp. |



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DEVOLUÇÃO
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para IDENTIFICAÇÃO
for IDENTIFICATION

DOAÇÃO
GIFT

Enviamos o material abaixo relacionado:
(We have sent the specimens listed below).

EMPRESTIMO por meses, renovável a pedido.
 LOAN for months, renewable on request.

Cerambycidae - Coleoptera
Ibidionini

1. *Notosphaeridion umbrinum* Martins 1 ex.-Holotype Manchere
2. *Tropidion hispidum* Martins 1 ex.-Holotype Manchere

38

Figures 34-38. 34-37) *Gnomibidion contiguum* n. sp., holotype ♂: 34) Dorsal habitus; 35) Ventral habitus; 36) Lateral habitus; 37) Head, frontal view. 38) Part of the invoice to SMFD.

Remarks

Neocompsa monnei n. sp. is similar to *N. ptoma* Martins, *Gnomibidion contiguum* n. sp. is similar to *G. biacutum* Martins, 1968 (see photograph of the holotype at Bezark 2019c), but differs by the prothorax not strongly constricted centrally, and elytral apex not noticeably acute, wider, with projection of outer angle shorter, and distinct projection at sutural angle. In *G. biacutum*, the prothorax is strongly constricted centrally, elytral apex is distinctly acute, narrower, with long projection at outer angle, and projection of sutural angle almost absent. It differs from *G. fulvipes* (Thomson, 1865) (see photographs at Bezark 2019c) by the prothorax lacking acute and distinct tubercles near middle (present in *G. fulvipes*); from *G. translucidum* (Martins, 1960) (see photographs at Bezark 2019c) by the prothorax not strongly constricted centrally (distinctly constricted in *G. translucidum*), prothorax lacking dark macula (present in *G. translucidum*), and spine of outer angle of the elytra distinctly shorter (long in *G. translucidum*). Finally, it differs from *G. occultum* Martins, 1968 by the elytra with distinct elliptical yellow macula margined with dark brown band anteriorly and posteriorly (absent in *G. occultum*).

Etymology. – The specific epithet *contiguum* refers to the large yellowish maculae that are contiguous along the elytral suture.

***Tropidion hispidum* Martins, 1971**

Tropidion hispidum Martins, 1971: 1441;
Joly, 1991: 29; Monné, 1993: 25 (cat.);
Monné & Giesbert, 1994: 73 (checklist);
Monné, 2005: 339 (cat.);
Monné & Hovore, 2006: 91 (checklist);
Martins & Galileo, 2007: 148;
Monné, 2019: 579 (cat.);
Bezark, 2019a: 139 (checklist).

According to the original description, the holotype belonged to “Natur-Museum und Forschungs-Institut Senckenberg.” [SMFD]. However, we found that the specimen was received by Elli Franz in 1972 (curator of SMFD at that time), and she sent the specimen to ZSMC (Fig. 38). Accordingly, the institution depository of the holotype of *Tropidion hispidum* is ZSMC, and not SMFD.

HEXOPLONINI Martins, 2006

According to Bouchard *et al.* (2011):

“Tribe Hexoplonini Martins, 2006. Hexoplonini Martins, 2006: 22 [stem : *Hexopl-*]. Type genus: *Hexoplon* J. Thomson, 1864. Comment: incorrect original stem formation, not in prevailing usage; this name was incorrectly treated as unavailable by Bousquet *et al.* (2009).”

The Code (ICZN 1999) does not establish how many works need to mention a name to be considered in “prevailing usage.” We believe that if there is no optional name, a single mention (the original one) must be considered as “prevailing usage:” **“usage, prevailing, n.** Of a name: that usage of the name which is adopted by at least a substantial majority of the most recent authors concerned with the relevant taxon, irrespective of how long ago their work was published.” Hexoplonini was used, for example, after the original description (and before Bouchard *et al.* 2011) by Monné & Hovore (2006), Clarke (2007), Martins *et al.* (2009a, b), Monné *et al.* (2009), Galileo & Martins (2009), Galileo & Martins (2010), Martins & Galileo (2010), Swift *et al.* (2010), Touroult *et al.* (2010), and Casari *et al.*

(2010). Thus, we think that the condition of the Article 29.5 of the ICZN (1999) is present (“Maintenance of current spellings.”). Therefore, we consider Hexoplonini Bouchard *et al.*, 2011 is an unjustified emendation.

***Hexoplon umbrinum* (Martins, 1971), comb. nov.**
(Figs. 39–42)

Notosphaeridion umbrinum Martins, 1971: 1421;
Joly, 1991: 10;
Monné, 1993: 4 (cat.);
Monné & Giesbert, 1994: 69 (checklist);
Monné, 2005: 320 (cat.);
Martins, 2006: 72;
Monné & Hovore, 2006: 87 (checklist);
Bezark, 2019a: 120 (checklist);
Monné, 2019: 452 (cat.).

Remarks

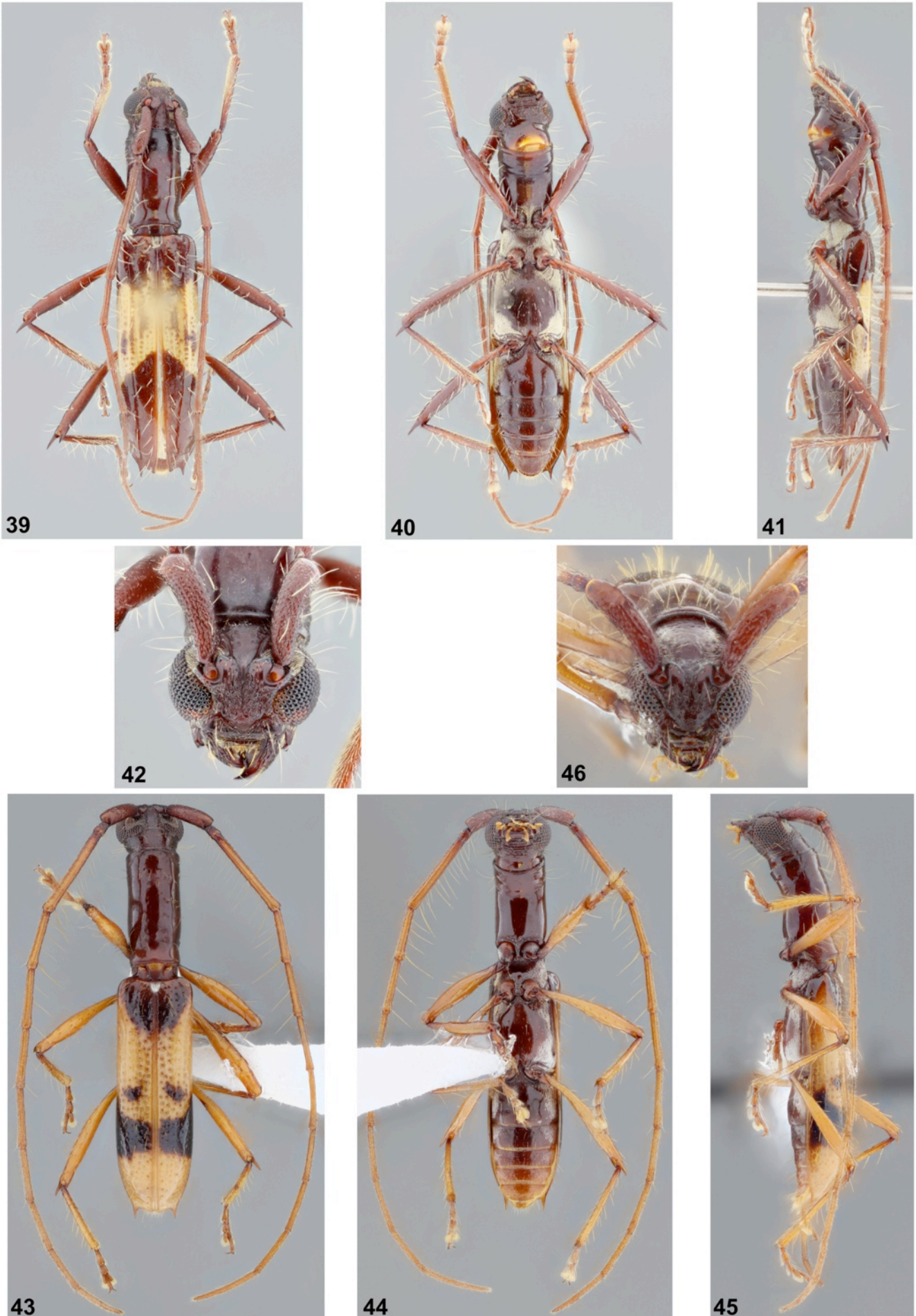
Martins (1960) described *Notosphaeridion* as follows (translated):

“*Antennal tubercles slightly projected; scape cylindrical, moderately elongated, strongly punctate; antennomere III bicarinate and the longest; antennomeres IV and V subequal in length, also bicarinate; prothorax slightly longer than wide, with distinct posterior constriction, globose centrally, and also with anterior constriction (less distinct than posterior one); pronotum shining, without tubercles; elytra coarsely punctate on anterior half; elytral apex with two subequal teeth in length; femora slender; mesofemora with short dentiform projection at inner apex; metafemora with two spiniform or dentiform projections; metatibiae carinate.*”

Three species were originally included in this genus: *N. scabrosum* (Gounelle, 1909), the type species; *N. brevithorax* (Martins, 1960); and *N. vestitum* Martins, 1960, new species described in the same work. These species agree well with the original description, and it is appropriate that all three are included in the genus.

Martins (1960) compared *Notosphaeridion* with *Gnomidolon* Thomson (1864) (translated): “By the shape of the femora and antennae, the genus, as stated above, approaches *Gnomidolon* Thomson. Moreover, Gounelle described the species we now choose for type species in *Gnomidolon*... *Notosphaeridion*, gen. n., differs from *Gnomidolon*, especially by the different prothoracic shape, which in this new genus is somewhat short, strongly constricted posteriorly, and rounded in the sides, while in *Gnomidolon* is elongated, cylindrical and without distinct constriction posteriorly. The punctuation and setae of the species included in the new genus are also distinctly different from that in *Gnomidolon*, in which the species are, often, barely punctate and setose. The elytral apex also offers a good differential character; in *Notosphaeridion*, gen. n., each elytral apex has two subequal spines, and in *Gnomidolon* they have only a spine of outer angle.” However, the prothoracic posterior constriction is also well-marked in several species of *Hexoplon* Thomson, 1864. Furthermore, currently, there are species in *Hexoplon* with the sides of the prothorax distinctly rounded between the anterior and posterior constrictions, and the inner spine of the apex of the mesofemora very similar to that of *Notosphaeridion brevithorax*.

A few years later, Martins (1967) included *Notosphaeridion* in his key to species of the Division I (currently, Hexoplonini) (translated): “12(11). Inner spine of apex of mesofemora well-developed, namely, very often as long as the outer spine of metafemora; metafemora may have another developed spine at



Figures 39-46. 39-42) *Hexoplon umbrinum*, ♀: 39) Dorsal habitus; 40) Ventral habitus; 41) Lateral habitus; 42) Head, frontal view. 43-46) *Gnomidolon sylvorum*, ♂: 43) Dorsal habitus; 44) Ventral habitus; 45) Lateral habitus; 46) Head, frontal view.

inner apex”, leading to *Hexoplon*; “Apex of mesofemora with only inner acute projection or with spine of short length, often shorter than outer spine of metafemora; metafemora with only spine at outer apex or with two short subequal projections” leading to *Notosphaeridion* and *Gnomidolon*. This key works well for the species included in *Notosphaeridion* by Martins (1960), but may lead to mistakes regarding some species of *Hexoplon* (including those known at that time).

Martins (1971) described the fourth species of *Notosphaeridion*: *N. umbrinum*, from Venezuela. This species disagrees with the original description of the genus in more than one feature: the prothorax is much longer than wider; the posterior constriction of the prothorax is not strong; the sides of the prothorax are not distinctly rounded; and the inner spine of the apex of mesofemora is as long as the outer spine of the metafemora. Following the definition of *Hexoplon* by Martins (1967), it is difficult to understand why *N. umbrinum* was not described in *Hexoplon*. Actually, Martins (1967) also questioned the separation between *Hexoplon* and *Gnomidolon* (this later compared with *Notosphaeridion* by him) (translated): “To separate *Hexoplon* from *Gnomidolon* the difficulties are greater. The only character that distinguishes them is the length of the inner spine of the apex of the mesofemora. This character is unsafe in some transitional species. We have seen above that previous authors, working on these genera, experienced analogous difficulty.”

Martins (2006) published a key to South American genera of Hexoplonini, and separated *Hexoplon*, *Notosphaeridion* and *Gnomidolon* in the alternative of couplet “18” (translated): “Inner spine of apex of mesofemora developed, often as long as outer spine of metafemora (in some species, the outer spine of the apex of mesofemora is long),” leading to *Hexoplon*; “Apex of mesofemora with inner projection just projected or with short spine (distinctly shorter than outer spine of apex of metafemora); metafemora with outer spine or inner and outer apex only projected,” leading to *Notosphaeridion* and *Gnomidolon*. However, as seen earlier, the inner spine of the apex of the mesofemora in *N. umbrinum* does not allow including the species in the second option of the alternative of couplet.

Joly (1991) already had indicated the proximity of *N. umbrinum* with *Hexoplon* (translated): “Mesofemora with inner spine very developed, as long as the outer spine of the metafemora, similar to that in the genus *Hexoplon*.”

Based on the impossibility of keeping *N. umbrinum* in *Notosphaeridion* by the absence of the features of the genus (shape of prothorax and mesofemora), it is transferred to *Hexoplon* where, we believe, it should have been originally described.

According to the original description, the holotype (the only known specimen at time of the original description) belonged to “Natur-Museum und Forshungs-Institut Senckenberg.” [SMFD]. However, according to Damir Kovac (SMFD, personal communication) the specimen is not there. Searching among the documents of specimens returned by Ubirajara R. Martins (in the archives of MZSP), we found that the specimen was received by Elli Franz in 1972 (curator of SMFD at that time), and she sent the specimen to ZSMC (Fig. 38). Accordingly, the institutional depository of the holotype of *Notosphaeridion umbrinum* is ZSMC, and not SMFD.

Material examined

– VENEZUELA, *Aragua*: Rancho Grande, 1100 m, 1 ♀, 23.V.1978, no collector indicated (MZSP).

Gnomidolon sylvarum (Bates, 1892)

(Figs. 43–46)

Hexoplon sylvarum Bates, 1892: 155;
Aurivillius, 1912: 105 (cat.);
Blackwelder, 1946: 568 (checklist).

Gnomidolon sylvarum;
Martins & Chemsak, 1966: 456;
Martins, 1967: 254; 1971: 128;
Chemsak *et al.*, 1992: 50 (checklist);
Monné, 1993: 16 (cat.);
Monné & Giesbert, 1994: 68 (checklist); Noguera & Chemsak, 1996: 399 (checklist);
Turnbow *et al.*, 2003: 12 (distr.);
Monné, 2005: 314 (cat.);
Monné & Hovore, 2006: 86 (checklist);
Hovore, 2006: 372 (distr.);
Monné, 2019: 444 (cat.);
Bezark, 2019a: 119 (checklist).

Remarks

Hexoplon sylvarum was described based on a single specimen from Mexico (Veracruz). Martins & Chemsak (1966) transferred this species to *Gnomidolon* Thomson, 1864 but did not report if they examined specimens. Martins (1967) examined and redescribed the holotype (the only specimen studied). Martins (1971) examined the second known specimen, recording the species from Guatemala, and reported (translated): “Specimen from Guatemala (HUB [Museum für Naturkunde, Humboldt-Universität, Berlin]) has the elytral drawing noticeably different from that of the holotype. In this specimen, the elytra have the base black in short extension, one small centrodorsal macula, triangular, also black, and a transverse ante-apical black macula. This pattern, with even greater reduction in basal and ante-apical bands, fits well with the description of *denticorne*; it is possible (as occurs with *Hexoplon albipenne* of similar pattern) that *sylvarum* and *denticorne* will be extreme forms of the same species. Examination of abundant material from Mexico and Central America will clarify the issue.”

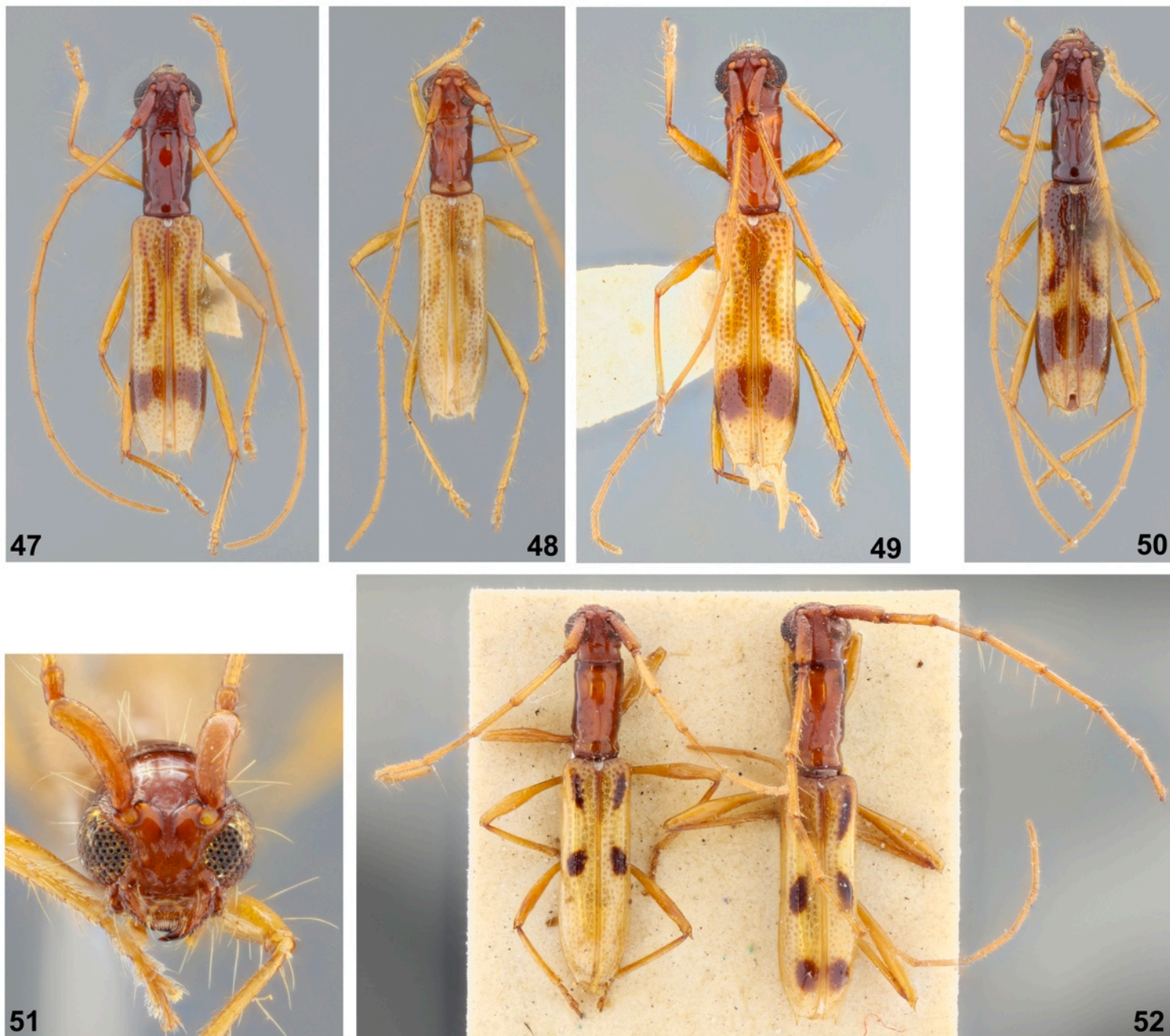
The specimen examined by us (Figs. 43–46) agrees very well with the specimen examined by Martins (1971) and, we believe it is *G. sylvarum*. We did not find any morphological difference to separate this specimen from the holotype of *G. sylvarum*, except the elytral color pattern, which we think is just variation. Furthermore, *G. varians* Gounelle, 1909 (Figs. 47–51) shows chromatic variations much more noticeably. This latter species differs from *G. sylvarum*, especially by the shorter prothorax (Figs. 47–50) (longer in *G. sylvarum* (Fig. 43)), and by the shining frons (Fig. 51) (opaque in *G. sylvarum* (Fig. 46)). This species is also very similar to *G. laetabile* Bates, 1885 (Fig. 52), but apparently it differs by the shorter spine of the outer femora, and the antennae are slightly shorter in males. Currently, this species is known from Mexico (Veracruz), Guatemala and Honduras (Monné 2019).

Material examined

– MEXICO, *Oaxaca* (New state record): 1–5 km N Hwy 200, road to San Antonio Cuixtla, 24–25.VI.2015, 1 ♂, J. Rifkind col. (LGBC).

Acknowledgments

We express our sincere thanks to Damir Kovac (SMFD) for the information on the holotypes of *Notosphaeridion umbrinum*. The first author thanks Jacques Rifkind for continually providing him with material from Mexico to examine, Kevin Williams who brought cerambycids collected in malaise traps by Frank Parker for study, and Rich Zack for material from his many Guatemala expeditions.



Figures 47-52. 47-51) *Gnomidolon varians*: 47) ♀ from Brazil (Bahia), dorsal habitus; 48) ♀ from Brazil (Minas Gerais), dorsal habitus; 49) ♀ from Brazil (Rio Grande do Sul), dorsal habitus; 50) ♀ from Brazil (São Paulo), dorsal habitus; 51) ♀ from Brazil (Minas Gerais), head, frontal view. 52) *Gnomidolon laetabile*, paralectotypes, ♂ (right), ♀ (left), dorsal habitus.

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Résumé

Bezark L. G. & Santos-Silva A., 2019. – Note sur les Hexoplonini et les Neoibidionini, avec la description de trois nouvelles espèces (Coleoptera, Cerambycidae, Cerambycinae). *Faunitaxys*, 7(17): 1 – 17.

Trois nouvelles espèces sont décrites: *Compsibidion tuberculatum* n. sp., du Costa Rica; *Neocompsa monnei* n. sp., du Mexique, et *Gnomibidion contiguum* n. sp., du Panama. Le mâle de *Neocompsa wappesi* Giesbert, 1998 est redécrit; première citation de l'espèce pour le Mexique. La variation chromatique dans *Heterachthes w-notatus* Linsley, 1935 est étudiée; première citation de l'espèce pour le Guatemala. Hexoplini Bouchard et al., 2011 est considéré comme un amendement injustifié pour Hexoplonini Martins, 2006. *Notosphaeridion umbrinum* Martins, 1971 est transféré dans le genre *Hexoplon* Thomson, 1864. La variation chromatique de *Gnomidolon sylvarum* (Bates, 1892) est rapportée; première citation de l'espèce pour le Mexique. L'institution dépositaire des holotypes de *Tropidion hispidum* Martins, 1971, et de *Notosphaeridion umbrinum* est corrigée.

Mots clés. – Coleoptera, Cerambycidae, Hexoplonini, Neoibidionini, *Neocompsa*, *Gnomibidion*, longicornes, taxonomie, Costa Rica, Mexique, Panama, Amérique Centrale.

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Faunitaxys

Volume 7, Numéro 17, Novembre 2019

SOMMAIRE

Note sur les Hexoplonini et les Neoibidionini, avec la description de trois nouvelles espèces (Coleoptera, Cerambycidae, Cerambycinae)

Larry G. Bezark & Antonio Santos-Silva 1 – 17

CONTENTS

New species and notes on Hexoplonini and Neoibidionini (Coleoptera, Cerambycidae, Cerambycinae)

Larry G. Bezark & Antonio Santos-Silva 1 – 17

Illustration de la couverture : Hotel Tirol, Encarnacion, Paraguay.

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Publié par l'Association Française de Cartographie de la Faune et de la Flore (AFCFF)